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## ORIGINAL ARTICLE

# The suboptimal fibrinolytic response in COVID-19 is dictated by high PAI-1

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## Abstract

**Background:** Severe COVID-19 disease is associated with thrombotic complications and extensive fibrin deposition. This study investigates whether the hemostatic complications in COVID-19 disease arise due to dysregulation of the fibrinolytic system.

**Methods:** This prospective study analyzed fibrinolytic profiles of 113 patients hospitalized with COVID-19 disease with 24 patients with non-COVID-19 respiratory infection and healthy controls. Antigens were quantified by Ella system or ELISA, clot lysis by turbidimetric assay, and plasminogen activator inhibitor-1 (PAI-1)/plasmin activity using chromogenic substrates. Clot structure was visualized by confocal microscopy.

**Results:** PAI-1 and its cofactor, vitronectin, are significantly elevated in patients with COVID-19 disease compared with those with non-COVID-19 respiratory infection and healthy control groups. Thrombin activatable fibrinolysis inhibitor and tissue plasminogen activator were elevated in patients with COVID-19 disease relative to healthy controls. PAI-1 and tissue plasminogen activator (tPA) were associated with more severe COVID-19 disease severity. Clots formed from COVID-19 plasma demonstrate an altered fibrin network, with attenuated fiber length and increased branching. Functional studies reveal that plasmin generation and clot lysis were markedly attenuated in COVID-19 disease, while PAI-1 activity was elevated. Clot lysis time significantly correlated with PAI-1 levels. Stratification of COVID-19 samples according to PAI-1 levels reveals significantly faster lysis when using the PAI-1 resistant (tPA) variant, tenecteplase, over alteplase lysis.

Claire S. Whyte and Megan Simpson contributed equally.

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**Conclusion:** This study shows that the suboptimal fibrinolytic response in COVID-19 disease is directly attributable to elevated levels of PAI-1, which attenuate plasmin generation. These data highlight the important prognostic potential of PAI-1 and the possibility of using pre-existing drugs, such as tenecteplase, to treat COVID-19 disease and potentially other respiratory diseases.

**KEYWORDS**

COVID-19, fibrin, fibrinolysis, PAI-1, vitronectin

## 1 | INTRODUCTION

Thrombosis is a common feature of severe COVID-19 disease arising from infection with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2).<sup>1,2</sup> Venous thromboembolism, particularly pulmonary embolism (PE),<sup>3,4</sup> but also arterial thromboembolic complications, are also prevalent.<sup>5</sup> Large vessel thrombi are present in almost half of patients who are critically ill with COVID-19 and focal platelet and fibrin microthrombi are described in over 80% of cases.<sup>6</sup> Importantly, these thromboembolic complications are observed despite prophylactic and full dose anticoagulation.<sup>4</sup>

The coagulopathy seen in COVID-19 disease is characterized by elevated fibrinogen and D-dimer.<sup>7</sup> However, unlike disseminated intravascular coagulation resulting from bacterial sepsis,<sup>8</sup> there is minimal impact on the APTT/PT time and very mild thrombocytopenia.<sup>7,9</sup> Nonetheless, these individuals are predisposed to excessive fibrin deposition, particularly within the pulmonary vasculature. Autopsy findings reveal evidence of thrombi in pulmonary arterioles and arteries with diffuse alveolar damage and hyaline membrane deposition,<sup>10</sup> even in patients with no clinical features of thromboembolism.<sup>11</sup> Dysregulation of hemostasis in patients with COVID-19 is closely connected to the inflammatory state, consequential to the cytokine storm and endotheliopathy induced by SARS-CoV-2 infection.<sup>12,13</sup> The natural anticoagulant properties of the endothelium, such as the expression of thrombomodulin (TM) and glycocalyx, are reduced and release of nitric oxide is impaired, increasing the hypercoagulability of the blood and promoting platelet and leucocyte adhesion.<sup>14,15</sup>

Platelets and neutrophils recruited to sites of endothelial damage become activated by multiple pathways. Neutrophils express ACE-2 and can be directly infected by SARS-CoV-2, resulting in chemokine release, complement system activation and platelet assisted neutrophil activation with formation of neutrophil extracellular traps (NETS). Prominent cytokines in the inflammatory 'storm' include interleukin-6 (IL-6), tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ) and C-reactive protein (CRP). IL-6 stimulates platelet production and activity, augments tissue factor expression on endothelial cells and monocytes, and promotes endothelial dysfunction.<sup>16,17</sup> Together, activated platelets, NETS, coagulation system dysregulation and endothelium disruption contribute to the prothrombotic state and resultant fibrin deposition.

### Essentials

- Severe COVID-19 disease is associated with thrombosis and fibrin deposition.
- This study investigates whether thrombotic complications arise due to dysregulation of the fibrinolysis.
- Plasminogen activator inhibitor-1 (PAI-1) and its stabilizing cofactor, vitronectin, are significantly elevated in patients with COVID-19 disease.
- Studies with tissue plasminogen activator variants reveal that PAI-1 drives hypofibrinolysis in COVID-19 disease.

D-dimer levels reflect both thrombin generation and proteolytic degradation of fibrin by plasmin. The high D-dimer levels in patients with COVID-19 indicate activation of the fibrinolytic system, but despite this thrombosis predominates, indicating that a suboptimal fibrinolytic response may underlie these prothrombotic changes in COVID-19.<sup>18</sup> Generation of plasmin is governed by the balance of the plasminogen activators, tissue plasminogen activator (tPA) and urokinase (uPA), and the main inhibitors of the system, plasminogen activator inhibitor-1 (PAI-1),  $\alpha_2$ -antiplasmin ( $\alpha_2$ AP) and the activated form of thrombin activatable fibrinolysis inhibitor (TAFI). Circulating PAI-1 exists in complex with the plasma glycoprotein, vitronectin, which acts to stabilize the inhibitor<sup>19,20</sup> and localize it within fibrin.<sup>21</sup> Levels of PAI-1 are low in the circulation, but it is released from the endothelium, circulating platelets and adipocytes following stimulation.<sup>22</sup> Studies have indicated that the balance of fibrinolytic activators and inhibitors are altered in COVID-19, however there is inadequate consensus on the drivers of the hypofibrinolytic state, with the suggestion that elevated fibrinogen,<sup>23,24</sup> TAFI,<sup>24,25</sup> tPA<sup>26,27</sup> and PAI-1<sup>24,28,29</sup> or decreased plasminogen<sup>28,30</sup> are responsible.

Herein, we investigate the fibrinolytic response in patients hospitalized with COVID-19 disease compared with non-COVID-19 respiratory infections and a healthy cohort. We determine that COVID-19 infection is associated with impaired plasmin generation and fibrinolysis, primarily due to elevated levels of PAI-1 which is associated with disease severity. Importantly, we show that strategies to overcome functional PAI-1 in COVID-19 disease may have

therapeutic potential to ameliorate persistence of fibrin and thrombosis in severely ill patients.

## 2 | METHODS

### 2.1 | Study design

A prospective observational cohort study was performed at Aberdeen Royal Infirmary and approved by Regional Ethics Committee (reference 16/NS/0055). Adult patients ( $\geq 18$  years) presenting between April 2020 and March 2021 with acute symptoms and signs of COVID-19, or who developed COVID-19 while in hospital, were included after obtaining informed consent to collect blood samples and access clinical data. Samples were taken as close to presentation as the consent process permitted and patients categorized as SARS-CoV-2 positive or negative on the basis of PCR testing. Patients were followed up until death or discharge and samples correlated to subsequent disease progression. Ethical approval to collect blood from 30 healthy volunteers was obtained from the University of Aberdeen College Ethical Review Board, CERB/2017/9/1411.

### 2.2 | Clinical data collection

For all patients, baseline premorbid demographics were extracted systematically, including, age, sex, body mass index (BMI), ethnicity, smoking status, known risk factors and medications. Dates of symptom onset and SARS-CoV-2 PCR test results were recorded along with presentation blood parameters taken for standard care. Data sources included patient paper and electronic hospital records, electronic hospital and primary care clinical summaries, TrakCare and patient reports. Consenting patients and/or their clinical records were reviewed daily during admission and an ordinal score representing the highest level of respiratory (and other organ) support recorded (Table 1). The maximum level of clinical support during hospital admission was grouped into mild—no oxygen support; moderate—oxygen delivered by face mask or normal nasal cannula; or severe—oxygen delivered through high flow nasal cannula (HFNC) or continuous positive airways pressure (CPAP) systems, invasive ventilation and above. Clinical notes and imaging results were reviewed for evidence of thromboembolism (venous or arterial) during admission, defined as high clinical suspicion (with or without bedside ultrasound evidence) or radiologically proven (consultant radiologist report).

### 2.3 | Laboratory sample preparation and handling

Blood samples were collected in 0.1 ml volume of 0.13M trisodium citrate. Platelet poor plasma was obtained by centrifugation of whole blood samples at  $2500 \times g$  for 30 min at 4°C. Pooled normal plasma (PNP) was prepared as described<sup>31</sup> or kindly provided by National Institute for Biological Standards and Control (NIBSC).

TABLE 1 Ordinal score of level of respiratory support

Ordinal scale	Level of respiratory support
1	No oxygen support
2	Oxygen support
3	HFNC
4	CPAP
5	Ventilated
6	Ventilated with MOF

Abbreviations: CPAP, continuous positive airways pressure; HFNC, high flow nasal cannula oxygen therapy; MOF, multiorgan failure.

### 2.4 | Determination of plasma protein concentrations

Concentrations of D-dimer, PAI-1, CRP, transforming growth factor  $\beta 1$  (TGF- $\beta 1$ ), IL-1 $\beta$ , IL-6, IL-8, TNF- $\alpha$ , uPA and leptin in plasma were determined using Simple Plex assays on Ella system, according to manufacturers' guidelines. Fibrinogen, tPA, plasminogen, TAFI, vitronectin concentrations and PAI-1 activity were quantified using ELISA from Molecular Innovations and soluble thrombomodulin (sTM) from R&D systems.

### 2.5 | Turbidity assays

Clots were formed in Tris buffered saline with Tween-20 (TBST; 10mM Tris, 140mM NaCl and 0.01% Tween-20 pH 7.4) from 30% PNP, patient or individual healthy volunteer plasma as described<sup>32</sup>  $\pm 300$  pM tPA (alteplase, Actilyse) or (tenecteplase, Metalyse). Absorbance readings at 405nm were taken every min for 8 h at 37°C on a Multiskan microplate photometer (Thermo Scientific).

### 2.6 | Plasmin generation assays

Plasmin generation in plasma (10%) was measured using 0.5mM S-2251 (Chromogenix)  $\pm 10$  nM tPA (alteplase) and Cyanogen bromide fibrinogen fragments (10  $\mu\text{g}/\text{ml}$ ; Technoclone). Absorbance readings at 405nm were taken every 30s for 8 h at 37°C on a Biotek Flx800 microplate reader.

### 2.7 | Confocal microscopy of fibrin structure

Plasma (30%) with 0.25  $\mu\text{M}$  AlexaFluor 488 (AF488) or 546 (AF546) fibrinogen in TBST,  $\text{CaCl}_2$  (10mM) and thrombin (0.125U/ml) was added to Ibidi VI 0.4 slides and the clots formed for 2 h at ambient temperature. Images were recorded on Zeiss 710 laser scanning confocal microscope with a 63 $\times$ 1.40 oil immersion objective using Zeiss Zen 2012 software.

TABLE 2 Patient demographics on presentation

	COVID-19 positive (113)	Non-COVID-19 respiratory infection (24)	p
Age, mean ( $\pm$ SD)	60.8 ( $\pm$ 12.6)	64.4 ( $\pm$ 18.3)	ns
Sex (%)			
Male	66 (58.4)	10 (41.7)	ns
Female	47 (41.6)	14 (58.3)	
BMI, mean ( $\pm$ SD)	31.9 <sup>a</sup> ( $\pm$ 6.3)	28.5 (6.9)	.02
Underweight < 18.5 (%)	0	1 (4.2)	
Normal 18.5–24.9 (%)	11 (9.7)	7 (29.2)	
Pre-obesity 25.0–29.9 (%)	32 (28.3)	5 (20.8)	
Obesity class I 30.0–34.9 (%)	35 (30.9)	5 (20.8)	
Obesity class II 35.0–39.9 (%)	24 (21.2)	5 (20.8)	
Obesity class III > 40 (%)	10 (8.8)	1 (4.2)	
Smoker (%)			
Yes	6 (5.3)	7 (29.2)	.002
At presentation			
Anticoagulant (%)	15 (13.3)	5 (20.8)	ns
Apixaban	2	3	
Fondaparinux	1	0	
Rivaroxaban	6	1	
Dabigatran	1	0	
Edoxaban	0	1	
Warfarin	5	0	
Antiplatelet (%)	24 (21.2)	6 <sup>a</sup> (25)	ns
Aspirin	14	3	
Clopidogrel	8	2	
Dual—aspirin and clopidogrel	2	0	
Dual—aspirin and ticagrelor	2	0	
ACE inhibitors/ARB (%)	36 (31.9)	4 (16.7)	ns
Medical history (%)			
DVT/PE	8 (7.1)	2 (8.3)	ns
CVA/TIA	8 (7.1)	1 (4.2)	ns
IHD	22 (19.5)	5 (20.8)	ns
PVD	5 (4.4)	1 (4.2)	ns
Heart failure	8 (7.1)	2 (8.3)	ns
Hypertension	36 (31.9)	7 (29.2)	ns
AF	0	0	ns
MI	0	0	ns
COPD/asthma	29 (25.6)	10 (41.7)	ns
Pulmonary other	9 (8.0)	2 (8.3)	ns
DM	28 (24.8)	5 (20.8)	ns
Immunosuppression	15 (13.3)	4 (16.7)	ns
CKD $\geq$ 3	10 (8.8)	4 (16.7)	ns
Liver disease	4 (3.5)	0	ns
Days hospitalized median (IQR)	7 (4.0–11)	3 (1.0–7.8)	<.001
Respiratory support (%)			
No oxygen support	16 (14.2)	13 (54.2)	<.001

(Continues)

TABLE 2 (Continued)

	COVID-19 positive (113)	Non-COVID-19 respiratory infection (24)	<i>p</i>
Oxygen support	70 (61.9)	10 (41.7)	ns
HFNC/CPAP	22 (19.45)	0	ns
Ventilated	5 (4.4)	0	ns
Thrombotic events	4	0	ns
Deaths (%)	18 (15.9)	2 (8.3)	ns
COVID-19 as cause of death (%)	17 (15.0)	n/a	

Note: Quantitative data were analyzed by an unpaired Student's *t*-test and expressed as mean  $\pm$  SD or analyzed with a Mann-Whitney test and expressed as median (IQR). Statistical significance for categorical data was analyzed using a Fisher's exact test.

Abbreviations: ACE, angiotensin-converting enzyme; AF, atrial fibrillation; ARBs, inhibitors and angiotensin II receptor blockers; BMI, body mass index; CKD, chronic kidney disease; COPD, chronic obstructive pulmonary disease; DM, diabetes mellitus; DVT, deep vein thrombosis; IHD, ischemic heart disease; MI, myocardial infarction; PE, pulmonary embolism; PVD, peripheral vascular disease.

<sup>a</sup>Indicates missing data for some patients.

## 2.8 | Western blotting

COVID-19 positive plasma samples and PNP were separated on 4–12% NuPAGE Bis-Tris gels under non-reducing conditions, then transferred to PVDF membrane. PAI-1 was detected with polyclonal PAI-1 antibody (Affinity Biologicals) and donkey anti-sheep IgG conjugated to HRP (Sigma).

## 2.9 | Statistical analysis

Statistical analysis was performed using GraphPad Prism Software (Version 9.1.0). Normally distributed data are expressed as mean  $\pm$  standard deviation and statistical significance tested using an unpaired Student's *t*-test. Otherwise, data are expressed as median  $\pm$  interquartile range (IQR) and either a Mann Whitney test or a Kruskal-Wallis test followed by a Dunn's multiple comparison test. Statistical significance for categorical data were analyzed using a Fisher's exact test. A value of *p* < .05 was considered statistically significant for all tests. Correlations were determined by Pearson correlation coefficients. Rate of plasmin generation was calculated using; Longstaff C, 2016, Shiny App for calculating zymogen activation rates, version 0.6 (<https://drclongstaff.shinyapps.io/zymogenactnCL/>). Confocal microscopy images were analyzed using ImageJ 1.51w and the Diameter J<sup>33</sup> plugin.

## 3 | RESULTS

Of the 137 patients presenting with suspected COVID-19 disease, 113 were positive by PCR. The remaining 24 tested negative and were managed for non-COVID-19 respiratory infection (Table S1). The baseline characteristics of the two groups were largely similar, although BMI was higher in the COVID-19 cohort (Table 2).

The median hospital stay of the COVID-19 cohort was significantly longer than those with non-COVID-19 respiratory infection

(7 [IQR 4–11] vs. 3 [IQR 1–7.8] days, Table 2) but the death rate, although higher (15.9% vs. 8.3%), did not reach significance. The mean age of the COVID-19 patients was 60.8  $\pm$  12.6 years, 66 (58.4%) were male and four (3.5%) experienced a thrombotic event (three PE and one ischemic stroke), of whom one required mechanical ventilation (Table 2). Presentation full blood counts (Table 3) showed significantly reduced leucocytes compared with patients with non-COVID-19 respiratory infections and reduced platelet counts, although these did not reach significance.

## 3.1 | Fibrinolytic antigens are increased in patients with COVID-19 and associate with clinical inflammatory biomarkers

We initially quantified several clinical diagnostic markers of inflammation and coagulation. CRP was significantly elevated in patients with COVID-19 compared with healthy controls (*p* < .001) and non-COVID-19 respiratory infection (*p* < .05, Figure 1A). Fibrinogen (*p* < .001, Figure 1B) and D-dimer (*p* < .001, Figure 1C) levels were raised in COVID-19 disease but only reached significance compared with healthy controls. In line with the well documented 'cytokine storm' in COVID-19, we observed significantly elevated levels of TGF- $\beta$ 1, IL-1 $\beta$ , IL-8, IL-6 and TNF- $\alpha$  (Figure 1D–H) compared with healthy controls. TNF- $\alpha$  was significantly higher in patients with COVID-19 compared with non-COVID-19 respiratory infection (*p* < .01, Figure 1G).

PAI-1 and its cofactor vitronectin were significantly elevated in patients with COVID-19 compared with healthy controls (*p* < .001) and non-COVID-19 respiratory infection (*p* < .01) (Figure 2A,B). We observed bands in COVID-19 plasma at a molecular mass consistent with the size of the reported PAI-1-Vn complex<sup>19</sup> (Figure S1). Of note, tPA and TAFI levels were elevated in patients with COVID-19 disease compared with healthy controls (*p* < .001 and *p* < .05, respectively) but were not different compared with non-COVID-19 respiratory infection (Figure 2C,F). Interestingly, plasminogen differed only between the two disease groups due to a reduced level in

TABLE 3 Presentation full blood counts

	COVID-19 positive (113)	Non-COVID-19 respiratory infection (24)	<i>p</i>
Hemoglobin (g/L)	140.0 (129.0–151.0)	130.5 (112.3–151.0)	ns
Platelets ( $\times 10^9$ /L)	213.5 (174.0–269.3)	243.0 (206.3–298.3)	ns
Leucocyte ( $\times 10^9$ /L)	6.6 (4.9–8.4)	12.2 (9.3–16.0)	<.001
Neutrophils ( $\times 10^9$ /L)	5.1 (3.5–6.5)	10.2 (7.0–13.8)	<.001
Lymphocytes ( $\times 10^9$ /L)	0.8 (0.6–1.3)	1.2 (7.0–13.8)	ns
Eosinophils ( $\times 10^9$ /L)	0.0 (0–0.02)	0.06 (0.0–0.18)	<.001
Basophils ( $\times 10^9$ /L)	0.0 (0–0.01)	0.02 (0.01–0.05)	.003
Monocytes ( $\times 10^9$ /L)	0.5 (0.3–0.6)	0.8 (0.4–1.2)	<.001

Note: Data are presented as median (IQR). Statistical difference was determined by Mann-Whitney test.

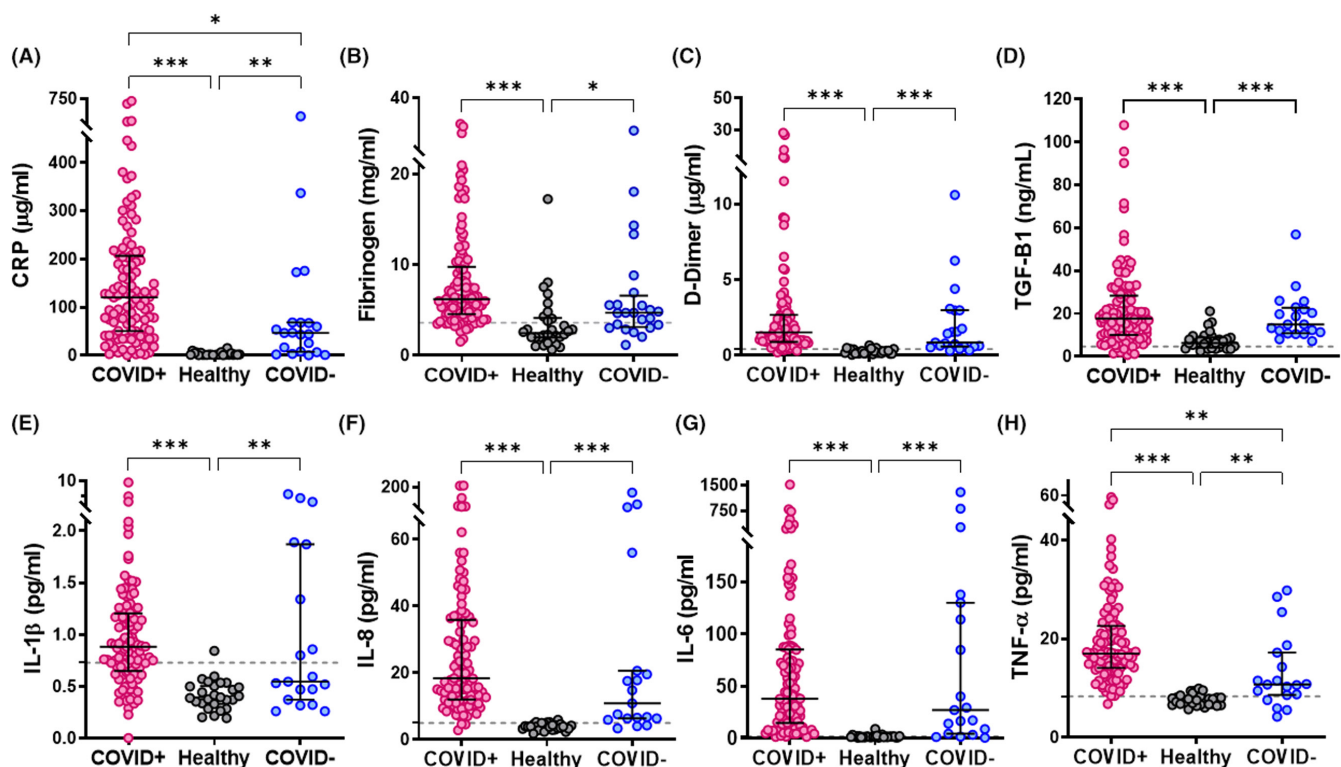


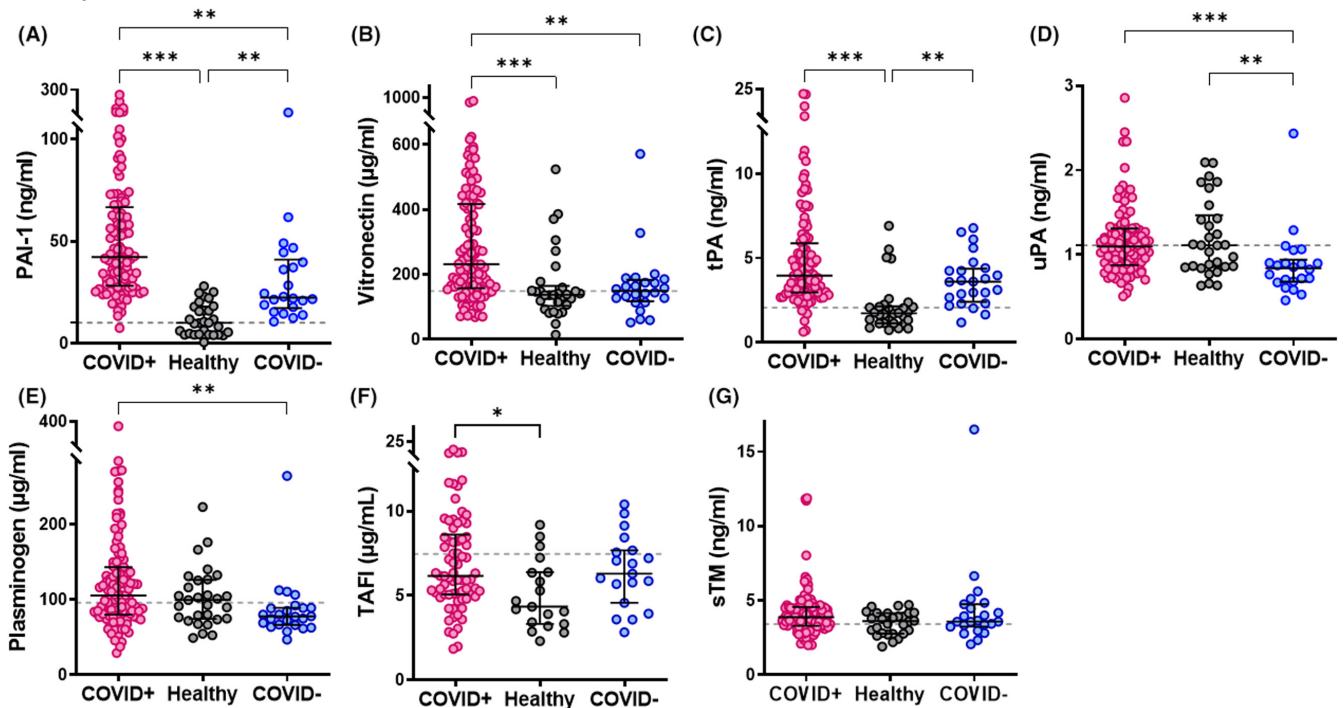
FIGURE 1 Inflammatory markers and proinflammatory cytokines in COVID-19. Antigen levels of inflammatory markers and proinflammatory cytokines were measured in plasma from confirmed COVID-19 patient plasma (COVID+), other non-COVID-19 respiratory infections (COVID-) or healthy controls by Simple Plex assays on Ella system or by ELISA for fibrinogen. (A) C-reactive protein (CRP), (B) fibrinogen, (C) D-dimer, (D) transforming growth factor  $\beta$ 1 (TGF- $\beta$ 1), (E) interleukin (IL)-1 $\beta$ , (F) IL-6, (G) IL-8 and (H) tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ). Dotted lines indicate antigen concentrations in pooled normal plasma. Data are shown as median  $\pm$  interquartile range (IQR), \**p* < .05, \*\**p* < .01 and \*\*\**p* < .001.

non-COVID respiratory disease (*p* < .01), with uPA also attenuated in this disease group (Figure 2D,E). There was no significant difference in sTM (Figure 2G).

The relationship between inflammatory and fibrinolytic markers with COVID-19 disease progression was studied by grouping according to the level of maximum oxygen support required. CRP and D-dimer increased with disease severity, with CRP being a

strong indicator of progression to more severe COVID-19 illness (Figure S2A,B). IL-6 and IL-8 also increased with disease progression (Figure S2D,E). PAI-1 and tPA were the only fibrinolytic proteins that stratified with escalating severity of COVID-19 (Figure S3A,D). Changes in PAI-1 activity did not reach statistical significance, potentially reflecting the difference in quantifying free versus total PAI-1.





**FIGURE 2** Fibrinolytic proteins are elevated in COVID-19. The concentrations of fibrinolytic proteins were measured in plasma from confirmed COVID-19 patient plasma (COVID+), other non-COVID-19 respiratory infections (COVID-) or healthy controls by Simple Plex assays on Ella system or by ELISA for plasminogen activator inhibitor-1 (PAI-1). (A) PAI-1, (B) vitronectin, (C) tissue plasminogen activator (tPA), (D) urokinase (uPA) (E) plasminogen, (F) thrombin activatable fibrinolysis inhibitor (TAFI) and (G) soluble thrombomodulin (sTM). Dotted lines indicate antigen concentrations in pooled normal plasma. Data are shown as median  $\pm$  interquartile range (IQR), \* $p < .05$ , \*\* $p < .01$  and \*\*\* $p < .001$ .

PAI-1 antigen was found to correlate strongly with TGF- $\beta$ 1 ( $r = .39$ ,  $p < .001$ ), IL-8 ( $r = .58$ ,  $p < .001$ ), moderately with IL-1 $\beta$  ( $r = .31$ ,  $p < .01$ ) and TNF- $\alpha$  ( $r = .33$ ,  $p < .001$ ). Additionally, a weak correlation was observed between PAI-1, fibrinogen ( $r = .27$ ,  $p < .01$ ) and D-dimer ( $r = .23$ ,  $p < .05$ ) but not with CRP (Figure S4). PAI-1 levels did not correlate with BMI in COVID-19 disease ( $r = .039$ ,  $p = .69$ ), despite the documented relationship with obesity. Leptin levels, which are known to correlate with body fat, were not different between the patient groups and there was no relationship with PAI-1 (Figure S5). Nonetheless, leptin significantly correlated with BMI in patients with COVID-19 (Figure S5).

### 3.2 | Suboptimal fibrinolytic potential and altered fibrin clot structure in COVID-19 disease

We then addressed the functional impact of changes in the balance of activators and inhibitors in COVID-19 disease. Elevated PAI-1 antigen correlated with an increase in PAI-1 activity ( $r = .55$ ,  $p < .001$ ), which was 3.8-fold higher in the plasma from patients with COVID-19 compared with healthy controls (Figure 3A,  $p < .001$ ). The rate of tPA-mediated plasmin generation (Figure 3B,  $p < .001$ ) was significantly attenuated compared with patients with non-COVID-19 disease and healthy controls, giving rise to slower rates of clot lysis (Figure 3C;  $p < .001$ ). A positive correlation existed between clot lysis

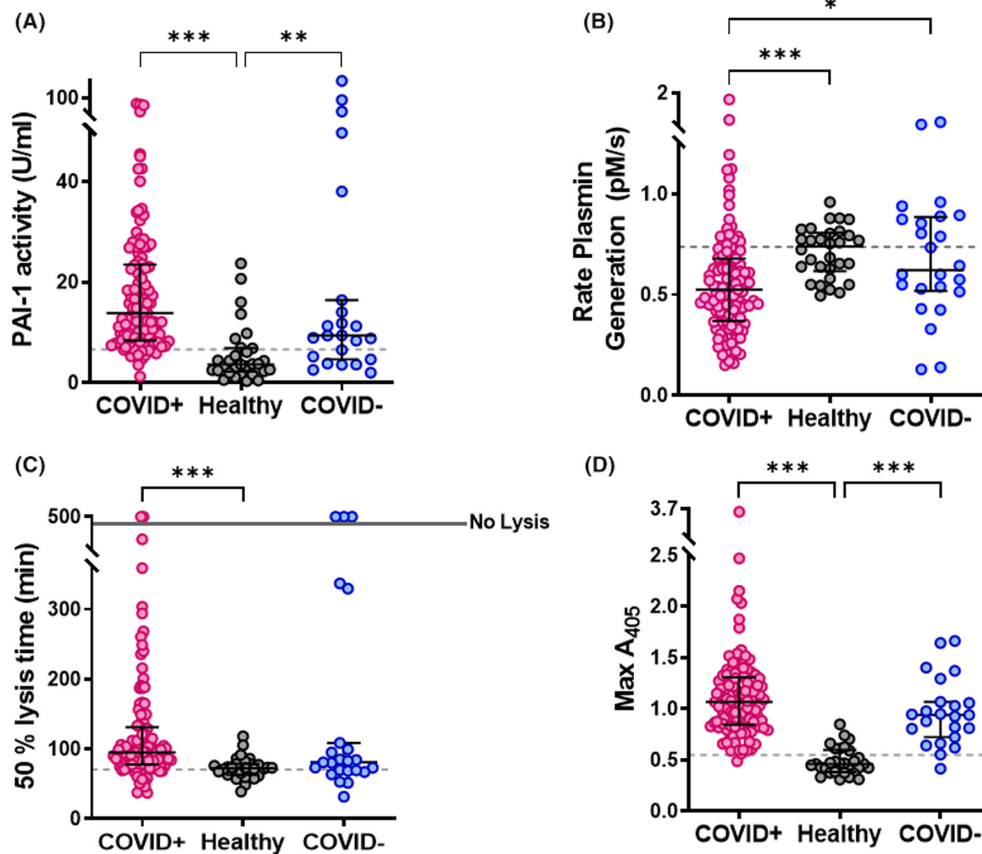
time and PAI-1 antigen ( $r = .3$ ,  $p = .001$ ) and activity ( $r = .74$ ,  $p < .001$ ) in patients with COVID-19 (Figure S6). Levels of fibrinogen and D-dimer also correlated with PAI-1 activity levels ( $r = .27$ ,  $p < .001$ ;  $r = .23$ ,  $p < .05$ , respectively; Figure S6).

The median maximum absorbance of plasma clots from the COVID-19 and non-COVID-19 respiratory infection was significantly higher than healthy controls (Figure 3D;  $p < .001$ ). Higher maximum absorbance can signify changes in fibrin clot structure and/or be reflective of baseline fibrinogen levels. Maximum absorbance correlated with fibrinogen ( $r = .29$ ,  $p < .01$ ; Figure S6) and vitronectin ( $r = .33$ ,  $p = .001$ ; Figure S7) in COVID-19 disease. Analysis of fibrin clot structure by confocal microscopy revealed a denser fibrin network with more branching, indicated by the increased number of intersections (Figure 4A,B,  $p < .05$ ) in patients who were COVID-19 positive compared with healthy controls. Fibrin fibers were also thinner (Figure 4A,C) and significantly shorter (Figure 4A,D,  $p < .01$ ).

### 3.3 | Elevated PAI-1 drives the hypofibrinolytic state in COVID-19 disease

To determine the direct contribution of PAI-1 in the reduced fibrinolytic potential in COVID-19 disease we utilized tenecteplase, a variant of tPA that is resistant to inhibition by PAI-1. Plasma clots



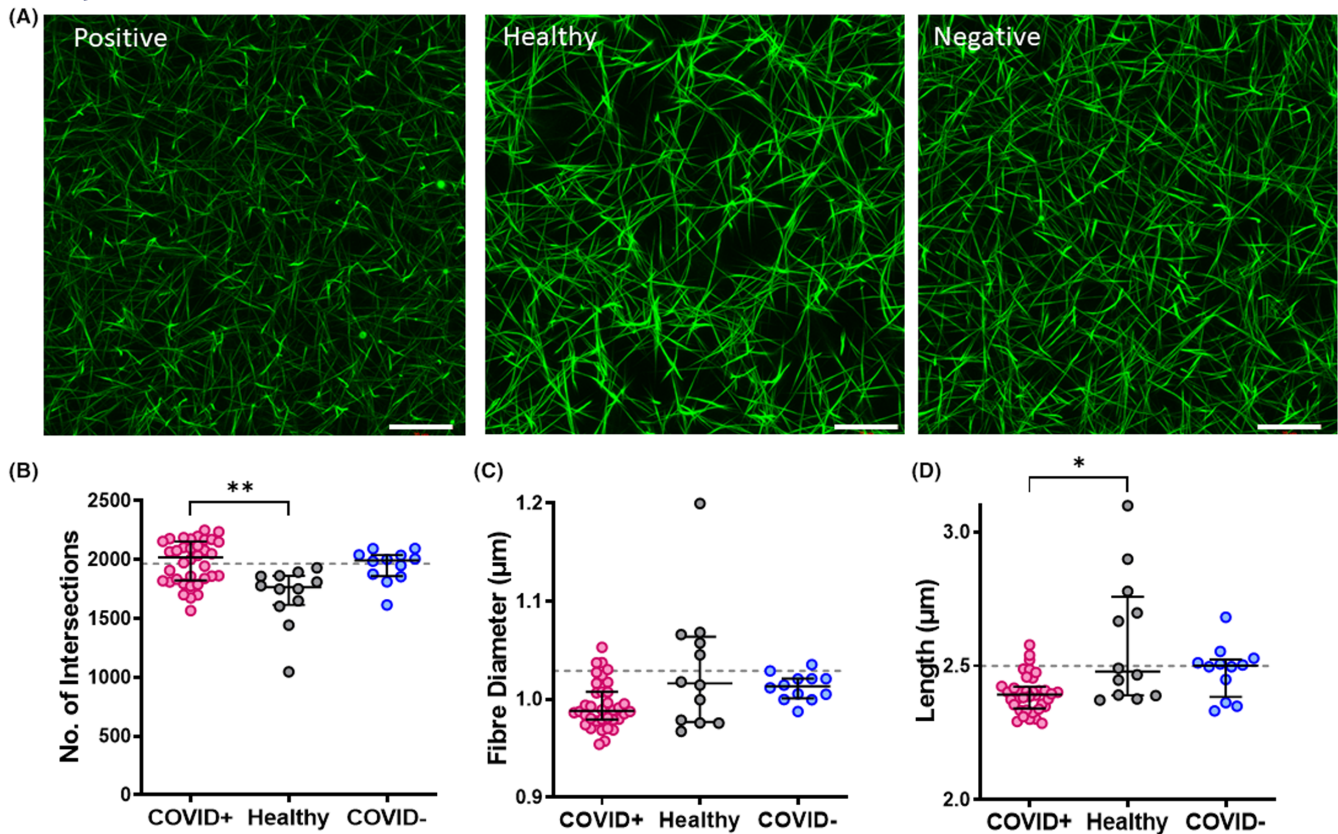


**FIGURE 3** COVID-19 disease is associated with reduced fibrinolytic potential. (A) Active plasminogen activator inhibitor-1 (PAI-1) was measured using a PAI-1 chromogenic activity assay. Data are presented as the median activity (U/ml)  $\pm$  IQR. (B) Plasmin generation in plasma (10%) was measured using S-2251 (0.5 mM) in the presence or absence of 10 nM alteplase with CBNR fibrinogen fragments (10  $\mu$ g/ml) included to stimulate tissue plasminogen activator (tPA) activity. The absorbance at 405 nm was read every 30 s for 8 h at 37°C. Data are the average rate of plasmin generation (pM/s)  $\pm$  IQR. (C) Plasma clots (30%) were formed with plasma from patients or healthy volunteers in the presence phospholipids (16 mM)  $\pm$  alteplase (300 pM). Clotting was initiated with CaCl<sub>2</sub> (10.6 mM) and thrombin (0.1 U/ml) and absorbance readings were taken every min for 8 h at 37°C. Data show the median  $\pm$  IQR time to 50% lysis and (D) maximum absorbance of plasma clots at 405 nm. \* $p$  < .05, \*\* $p$  < .01 and \*\*\* $p$  < .001.

were initially formed with PNP with various amounts of purified PAI-1 and lysed with alteplase versus tenecteplase. A dose-dependent increase in clot lysis time was observed with alteplase on addition of PAI-1, however lysis by tenecteplase was unaffected by addition of the inhibitor (Figure S8A). A subset of the COVID-19 samples were stratified according to their PAI-1 antigen levels (low <50 ng/ml [13.4–46.4 ng/ml]; medium 50–100 ng/ml [51.5–92.3 ng/ml]; high >100 ng/ml [103.3–260 ng/ml]) and clots lysis studies performed with alteplase and tenecteplase. At low PAI-1 concentrations, clots from patients with COVID-19 lysed at similar rates with alteplase and tenecteplase (Figure 5A & Figure S8B). In contrast, plasma clots with high PAI-1 lysed significantly slower with alteplase compared with tenecteplase (Figure 5A & Figure S8B). A strong correlation between 50% lysis time and PAI-1 concentration existed with alteplase ( $r = .68$ ,  $p < .001$ ), which was reduced with tenecteplase ( $r = .45$ ,  $p < .05$ , Figure 5B). No relationship was observed between 50% lysis time and endogenous tPA antigen (Figure 5C). These data suggest that there is a shift in the balance of fibrinolysis in COVID-19 disease towards inhibition, arising from high PAI-1.

## 4 | DISCUSSION

We examined the hemostatic dysregulation, specifically the contribution of the fibrinolytic pathway, in a large prospective cohort of patients hospitalized with COVID-19 disease. Our findings reveal marked elevation of PAI-1 and its stabilizing cofactor vitronectin in COVID-19 disease compared with non-COVID-19 respiratory infections. High PAI-1 suppressed plasmin generation and fibrinolysis in COVID-19, despite concomitant increases in tPA. Stratification of COVID-19 plasma according to PAI-1 levels revealed delayed lysis with alteplase with increasing PAI-1 but no difference in tenecteplase lysis. These functional data demonstrate that it is the high PAI-1 in COVID-19 disease that dictates the suboptimal fibrinolytic response. PAI-1 levels were associated with several inflammatory cytokines known to be elevated in COVID-19 disease, including IL-8, IL-6, TGF- $\beta$ 1 and TNF- $\alpha$ . Interestingly, higher levels of PAI-1 were associated with a poorer clinical outcome, suggesting that it may be a useful therapeutic target and/or a potential biomarker to help stratify those at risk of developing more severe COVID-19.

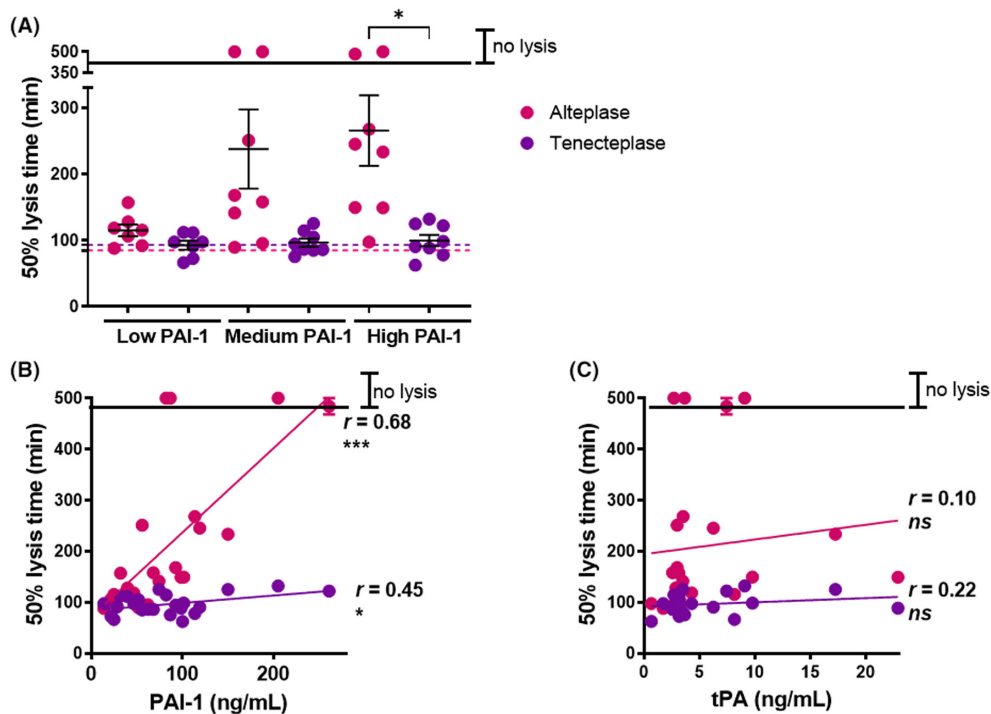


**FIGURE 4** Clots formed from COVID-19 plasma exhibit a denser fibrin network. Plasma clots (30%) were formed in the presence of AF488 or AF546 labeled fibrinogen (0.25  $\mu\text{M}$ ) by initiating clotting with  $\text{CaCl}_2$  (10 mM) and thrombin (0.125 U/ml) in Ibidi VI 0.4  $\mu$  slides. After allowing to clot for 1 h images were recorded on Zeiss 710 laser scanning confocal microscope with a  $63\times 1.40$  oil immersion objective using Zeiss Zen 2012 software. (A) Representative images of clots formed from plasma of a patient who was COVID-19 positive (COVID+), healthy donor and non-COVID-19 respiratory infections (COVID-). (B) Quantification of the number of intersections, which is indicative of branching and clot density. (C) Quantification of fiber diameter (D) Quantification of mean fibrin fiber length. Representative images of  $n \geq 3$ . Scale bar indicates 20  $\mu\text{m}$ . \* $p < .05$  and \*\* $p < .01$ .

Plasminogen activators circulate at low concentrations, while PAI-1 and  $\alpha_2\text{AP}$  exist in molar excess, preventing unrestricted plasmin activity. The primary source of tPA in the circulation is vascular endothelial cells, with release in response to specific stimuli, including bradykinin and  $\text{TNF-}\alpha$ . Other hemostatic proteins derived from the endothelium, including P-selectin, vWF and factor VIII, are upregulated in COVID-19 disease.<sup>34</sup> We found significantly elevated levels of tPA in patients with COVID-19 compared to healthy controls, but not to other non-COVID-19 respiratory infections. However, the levels reported herein are several fold lower than those described by Zuo and colleagues,<sup>26</sup> perhaps reflecting differences in methodology and/or patient cohort. We observed an association between increasing levels of tPA and COVID-19 disease severity, similar to observations in a smaller cohort.<sup>35</sup> Interestingly, plasminogen and uPA levels were not altered in COVID-19 disease but were reduced in non-COVID-19 respiratory infections. A longitudinal study of 14 patients with COVID-19 indicated a reduction in plasminogen with increasing COVID-19 disease severity,<sup>28</sup> but we did not observe significant differences in plasminogen in this large cohort of patients stratified according to severity.

Fibrinogen and thrombin concentration directly impact the quality of the fibrin network, with dense networks composed of thin fibers being directly associated with the pathophysiology of arterial and venous thromboembolic complications.<sup>36</sup> We describe elevated levels of fibrinogen in COVID-19 disease, as previously reported.<sup>23,24,37</sup> Similar to a recent publication, we found evidence of a dense fibrin network composed of thinner and shorter fibers with increased numbers of branching points.<sup>37</sup> Given the clear relationship between compact clot structure and down-regulation of fibrinolysis,<sup>36</sup> abnormal structure may contribute to persistence of fibrin within the microenvironment of the lung and systemically in critically ill individuals with COVID-19 disease.

Our work demonstrates a marked decrease in fibrinolytic potential in COVID-19 disease, observed as a reduction in plasmin generation and fibrinolysis, despite elevated tPA. These results contrast a previous report comparing COVID-19 disease and sepsis, which showed increased endogenous plasmin potential in patients with COVID-19.<sup>38</sup> However, this group also reported an increase in PAI-1 activity in COVID-19, in line with our observations.<sup>39</sup> Elevated levels of the fibrinolytic inhibitors, PAI-1 and TAFI in COVID-19 disease



**FIGURE 5** The tissue plasminogen activator (tPA) variant, tenecteplase, overcomes the reduced fibrinolytic potential in COVID-19 plasma. (A) Plasma clots (30%) were formed with COVID-19 plasma with low <50 ng/ml (13.4–46.4 ng/ml); medium 50–100 ng/ml (51.5–92.3 ng/ml) or high >100 ng/ml (103.3–260 ng/ml) PAI-1 antigen in the presence of phospholipids (16 mM) ± alteplase or tenecteplase (300 pM). Clotting was initiated with  $\text{CaCl}_2$  (10.6 mM) and thrombin (0.1 U/ml) and absorbance readings were taken every min for 8 h at 37°C. Data show the median ± IQR time to 50% lysis of plasma clots at 405 nm. (B) Correlation of 50% lysis time with PAI-1 antigen (C) or tPA antigen. \* $p < .05$  and \*\*\* $p < .001$ .

may promote the suboptimal fibrinolytic response in these individuals. TAFI was elevated in our cohort of patients with COVID-19 compared with healthy individuals, yet there was not a concomitant increase in its cofactor, TM. These observations were surprising, given other reports on increased levels of sTM due to endothelialopathy in COVID-19 disease.<sup>40</sup> In contrast, we show marked elevation of PAI-1 and its cofactor vitronectin in COVID-19 disease. Vitronectin serves not only to extend the half-life of circulating PAI-1<sup>19,41</sup> but importantly localizes this serpin within the fibrin network.<sup>21</sup> It is this latter aspect that is suspected to confer the enhanced fibrinolytic resistance as PAI-1 inhibition of fibrin clot lysis is dependent on the concentration of vitronectin.

To delineate the role of PAI-1 in driving the suboptimal fibrinolytic state in COVID-19 disease we compared lysis of plasma clots from patients with COVID-19 stratified according to PAI-1 levels. A direct correlation was observed between alteplase lysis time and PAI-1 concentration in COVID-19 plasma. Use of tenecteplase, a variant of alteplase with an 80-fold reduction in sensitivity to PAI-1,<sup>42</sup> was able to restore the fibrinolytic potential in plasma from patients with COVID-19 with high PAI-1 levels. These data directly attribute the suboptimal fibrinolytic response in patients with COVID-19 to PAI-1 that is likely sustained by elevated vitronectin. Therapeutic use of alteplase has gained interest as an intervention to target fibrin deposition in patients with COVID-19.<sup>43,44</sup> Our data show the superiority of tenecteplase to overcome PAI-1 in COVID-19 disease

and suggest that the clinical use of this variant as a thrombolytic may have improved benefit over alteplase.

Plasma PAI-1 can be derived from platelets, the vascular endothelium and adipocytes. The source of plasma PAI-1 in COVID-19 disease is unclear, but it has been suggested to relate to increased obesity in severely ill patients.<sup>18</sup> BMI was significantly higher in our cohort of patients with COVID-19 compared with those with non-COVID-19 respiratory illness but nonetheless there was no relationship between PAI-1 and BMI or with leptin, a biomarker of metabolic syndrome.<sup>45</sup> In contrast, leptin significantly correlated with increasing BMI in the COVID-19 cohort. These data suggest adipocytes may not be the source of PAI-1. Platelets contain the major pool of circulating PAI-1, which is released upon activation into the surrounding milieu and retained on the surface of activated platelets.<sup>46,47</sup> Our previous data have shown that vitronectin is released with PAI-1 from activated platelets.<sup>47</sup> Markers of ongoing platelet activation,<sup>48</sup> including P-selectin,<sup>49</sup> are well documented in patients with COVID-19, therefore it is plausible that degranulation of platelets could account for these increases in the plasma pool of PAI-1 and vitronectin. Activation of the endothelium and release of PAI-1 in response to cytokine and thrombin stimulation may also promote increased systemic levels of PAI-1. Interestingly, both PAI-1 and vitronectin were previously found to be elevated in SARS-CoV, a coronavirus which emerged almost 20 years prior

in 2002.<sup>50</sup> Regardless of the source of increased levels of these proteins, these data indicate that the PAI-1-vitronectin axis may be of importance in contributing to fibrin persistence and lung damage and thereby warrants attention as a potential therapeutic target.

Fibrinolytic and proinflammatory markers including D-dimer, CRP and fibrinogen were significantly elevated in this cohort of hospitalized patients with COVID-19, consistent with previous reports.<sup>7,51-55</sup> Notably, only CRP reached statistical significance in patients with COVID-19 compared with non-COVID-19 respiratory infection. CRP has been shown to provoke local release of PAI-1 from endothelial cells<sup>56,57</sup> thereby fostering further fibrin persistence in the alveolar space. In addition, several proinflammatory cytokines promote PAI-1 synthesis. IL-8, which correlated strongly with PAI-1 in this cohort, perturbs the balance of PAI-1 and tPA in endothelial cells<sup>58</sup> and may foster the high systemic PAI-1 levels. Models of SARS-CoV infection have demonstrated increased TGF- $\beta$ 1, which in turn induces PAI-1,<sup>59</sup> which is linked to development of ARDS.<sup>60</sup> We found elevated TGF- $\beta$ 1 in severe COVID-19 disease, which strongly correlates with PAI-1. A non-biased systems biology approach to study infection with SARS-CoV revealed over-expression of PAI-1 leading to fibrin persistence.<sup>61</sup> Furthermore, IL-6, an acute-phase protein that stimulates synthesis of both PAI-1 and tPA,<sup>62</sup> is upregulated in this cohort. Interestingly, tocilizumab, an IL-6 antagonist, which at has been used successfully to combat severe COVID-19 disease,<sup>63</sup> reduced PAI-1 in this group of patients providing direct evidence of the link between the proinflammatory state and PAI-1 levels.<sup>64</sup> The endotheliopathy and prothrombotic state are known to persist in patients with COVID-19 4–12 months post-discharge,<sup>18,65</sup> with elevated PAI-1 a component of this syndrome. It remains to be seen whether therapeutic agents, like tocilizumab, that dampen the inflammatory response, could be beneficial in reducing the prothrombotic tendency of these individuals.

#### 4.1 | Limitations

The unprecedented nature of the COVID-19 pandemic led to practical difficulties in taking consented research samples, however most samples were taken within 24 h of presentation or diagnosis. Administration of thromboprophylaxis is routine in hospital inpatients and those in critical care environments may have received higher prophylactic doses, mainly of low molecular weight heparin. Thrombosis outcomes were based upon attending clinical opinion and/or from radiological report. These interventions may have had a bearing on the reduced rates of death and thrombotic complications and levels of D-dimer observed in the presented cohort compared with earlier studies, such as Tang et al.<sup>7</sup> In addition, during the time frame of this study new treatment options, such as dexamethasone and tocilizumab, were introduced; however, as patients were sampled within 24 h these are unlikely to have bearing on the results.

## 5 | CONCLUSIONS

Our data demonstrate that PAI-1 is systemically elevated in COVID-19 disease, and that there is a concomitant increase in its co-factor vitronectin which confers fibrinolytic resistance. Importantly, our functional studies, utilizing PAI-1 resistant forms of tPA, reveal the pivotal role of PAI-1 in dictating the suboptimal fibrinolytic response during SARS-CoV2 infection. The reduction in fibrinolytic potential occurs despite elevated tPA, underscoring the dominant role of PAI-1 in this setting. Our data accentuate the prognostic value of PAI-1 and the potential to repurpose existing drugs, such as tenecteplase, to treat the hypofibrinolytic state in COVID-19 and other acute respiratory diseases.

#### AUTHOR CONTRIBUTIONS

CSW, MS and GBM designed, executed, analyzed research and wrote/edited the manuscript. CAW executed and analyzed the research. AJM, JCK, SS, NC and CNB collected samples and reviewed the manuscript. HW and JGC collected samples, analyzed clinical and laboratory data, wrote and edited the manuscript. NJM designed, executed, analyzed, managed the research and wrote/edited the manuscript.

#### CONSENT STATEMENT

All authors provide consent to publication of this data.

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#### CONFLICT OF INTEREST

The authors have no relevant conflict of interests to declare.



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## REFERENCES

- Bompard F, Monnier H, Saab I, et al. Pulmonary embolism in patients with COVID-19 pneumonia. *Eur Respir J*. 2020;56(1):2001365.
- Lodigiani C, Iapichino G, Carenzo L, et al. Venous and arterial thromboembolic complications in COVID-19 patients admitted to an academic hospital in Milan, Italy. *Thromb Res*. 2020;191:9-14.
- Klok FA, Kruip M, van der Meer NJM, et al. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thromb Res*. 2020;191:145-147.
- Litjens JF, Leclerc M, Chochois C, et al. High incidence of venous thromboembolic events in anticoagulated severe COVID-19 patients. *J Thromb Haemost*. 2020;18(7):1743-1746.
- Kunutsor SK, Laukkanen JA. Incidence of venous and arterial thromboembolic complications in COVID-19: A systematic review and meta-analysis. *Thromb Res*. 2020;196:27-30.
- Borczuk AC, Salvatore SP, Seshan SV, et al. COVID-19 pulmonary pathology: a multi-institutional autopsy cohort from Italy and New York City. *Mod Pathol*. 2020;33(11):2156-2168.
- Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost*. 2020;18(4):844-847.
- Iba T, Levy JH, Warkentin TE, et al. Diagnosis and management of sepsis-induced coagulopathy and disseminated intravascular coagulation. *J Thromb Haemost*. 2019;17(11):1989-1994.
- Langer F, Kluge S, Klamroth R, Oldenburg J. Coagulopathy in COVID-19 and its implication for safe and efficacious thromboprophylaxis. *Hamostaseologie*. 2020;40(3):264-269.
- Dolhnikoff M, Duarte-Neto AN, de Almeida Monteiro RA, et al. Pathological evidence of pulmonary thrombotic phenomena in severe COVID-19. *J Thromb Haemost*. 2020;18(6):1517-1519.
- Lax SF, Skok K, Zechner P, et al. Pulmonary arterial thrombosis in COVID-19 with fatal outcome: results from a prospective, single-center, clinicopathologic case series. *Ann Intern Med*. 2020;173(5):350-361.
- Iba T, Connors JM, Levy JH. The coagulopathy, endotheliopathy, and vasculitis of COVID-19. *Inflamm Res*. 2020;69(12):1181-1189.
- Bonaventura A, Vecchie A, Dagna L, et al. Endothelial dysfunction and immunothrombosis as key pathogenic mechanisms in COVID-19. *Nat Rev Immunol*. 2021;21(5):319-329.
- Mancini I, Baronciani L, Artoni A, et al. The ADAMTS13-von Willebrand factor axis in COVID-19 patients. *J Thromb Haemost*. 2021;19(2):513-521.
- Bazzan M, Montaruli B, Sciascia S, Cosseddu D, Norbiato C, Roccatello D. Low ADAMTS 13 plasma levels are predictors of mortality in COVID-19 patients. *Intern Emerg Med*. 2020;15(5):861-863.
- Du FH, Liu B, Zhang SY. COVID-19: the role of excessive cytokine release and potential ACE2 down-regulation in promoting hypercoagulable state associated with severe illness. *J Thromb Thrombolys*. 2021;51(2):313-329.
- Wang J, Jiang M, Chen X, Montaner LJ. Cytokine storm and leukocyte changes in mild versus severe SARS-CoV-2 infection: review of 3939 COVID-19 patients in China and emerging pathogenesis and therapy concepts. *J Leukoc Biol*. 2020;108(1):17-41.
- von Meijenfeldt FA, Havervall S, Adelmeijer J, et al. Prothrombotic changes in patients with COVID-19 are associated with disease severity and mortality. *Res Pract Thromb Haemost*. 2021;5(1):132-141.
- Declerck PJ, De Mol M, Alessi MC, et al. Purification and characterization of a plasminogen activator inhibitor 1 binding protein from human plasma. Identification as a multimeric form of S protein (vitronectin). *J Biol Chem*. 1988;263(30):15454-15461.
- Lawrence DA, Berkenpas MB, Palaniappan S, Ginsburg D. Localization of vitronectin binding domain in plasminogen activator inhibitor-1. *J Biol Chem*. 1994;269(21):15223-15228.
- Podor TJ, Peterson CB, Lawrence DA, et al. Type 1 plasminogen activator inhibitor binds to fibrin via vitronectin. *J Biol Chem*. 2000;275(26):19788-19794.
- Morrow GB, Whyte CS, Mutch NJ. A serpin with a finger in many PAIs: PAI-1's central function in thromboinflammation and cardiovascular disease. *Front Cardiovasc Med*. 2021;8:653655.
- Schrick D, Tokes-Fuzesi M, Reger B, Molnar T. Plasma fibrinogen independently predicts hypofibrinolysis in severe COVID-19. *Metabolites*. 2021;11(12):826.
- Bachler M, Bosch J, Sturzel DP, et al. Impaired fibrinolysis in critically ill COVID-19 patients. *Br J Anaesth*. 2021;126(3):590-598.
- Claesen K, Sim Y, Bracke A, et al. Activation of the carboxypeptidase U (CPU, TAFIa, CPB2) system in patients with SARS-CoV-2 infection could contribute to COVID-19 hypofibrinolytic state and disease severity prognosis. *J Clin Med*. 2022;11(6):1494.
- Zuo Y, Warnock M, Harbaugh A, et al. Plasma tissue plasminogen activator and plasminogen activator inhibitor-1 in hospitalized COVID-19 patients. *Sci Rep*. 2021;11(1):1580.
- Douin DJ, Shaefi S, Brenner SK, et al. Tissue plasminogen activator in critically ill adults with COVID-19. *Ann Am Thorac Soc*. 2021;18(11):1917-1921.
- Juneja GK, Castelo M, Yeh CH, et al. Biomarkers of coagulation, endothelial function, and fibrinolysis in critically ill patients with COVID-19: a single-center prospective longitudinal study. *J Thromb Haemost*. 2021;19(6):1546-1557.
- Nougier C, Benoit R, Simon M, et al. Hypofibrinolytic state and high thrombin generation may play a major role in SARS-COV2 associated thrombosis. *J Thromb Haemost*. 2020;18(9):2215-2219.
- Della-Morte D, Pacifici F, Ricordi C, et al. Low level of plasminogen increases risk for mortality in COVID-19 patients. *Cell Death Dis*. 2021;12(8):773.
- Mutch NJ, Moore NR, Wang E, Booth NA. Thrombus lysis by uPA, scuPA and tPA is regulated by plasma TAFI. *J Thromb Haemost*. 2003;1(9):2000-2007.
- Westbury SK, Whyte CS, Stephens J, et al. A new pedigree with thrombomodulin-associated coagulopathy in which delayed fibrinolysis is partially attenuated by co-inherited TAFI deficiency. *J Thromb Haemost*. 2020;18(9):2209-2214.
- Hotaling NA, Bharti K, Kriel H, Simon CG Jr. DiameterJ: a validated open source nanofiber diameter measurement tool. *Biomaterials*. 2015;61:327-338.
- Devreese KMJ. COVID-19-related laboratory coagulation findings. *Int J Lab Hematol*. 2021;43(Suppl 1):36-42.
- Henry BM, Cheruiyot I, Benoit JL, et al. Circulating levels of tissue plasminogen activator and plasminogen activator inhibitor-1 are independent predictors of coronavirus disease 2019 severity: a prospective, observational study. *Semin Thromb Hemost*. 2021;47(4):451-455.

36. Undas A, Ariens RA. Fibrin clot structure and function: a role in the pathophysiology of arterial and venous thromboembolic diseases. *Arterioscler Thromb Vasc Biol.* 2011;31(12):e88-e99.
37. Wygrecka M, Birnhuber A, Seeliger B, et al. Altered fibrin clot structure and dysregulated fibrinolysis contribute to thrombosis risk in severe COVID-19. *Blood Adv.* 2022;6(3):1074-1087.
38. Bouck EG, Denorme F, Holle LA, et al. COVID-19 and sepsis are associated with different abnormalities in plasma procoagulant and fibrinolytic activity. *Arterioscler Thromb Vasc Biol.* 2021;41(1):401-414.
39. Campbell RA, Hisada Y, Denorme F, et al. Comparison of the coagulopathies associated with COVID-19 and sepsis. *Res Pract Thromb Haemost.* 2021;5(4):e12525.
40. Goshua G, Pine AB, Meizlish ML, et al. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. *Lancet Haematol.* 2020;7(8):e575-e582.
41. Lawrence DA, Palaniappan S, Stefansson S, et al. Characterization of the binding of different conformational forms of plasminogen activator inhibitor-1 to vitronectin. Implications for the regulation of pericellular proteolysis. *J Biol Chem.* 1997;272(12):7676-7680.
42. Nordt TK, Bode C. Thrombolysis: newer thrombolytic agents and their role in clinical medicine. *Heart.* 2003;89(11):1358-1362.
43. Whyte CS, Morrow GB, Mitchell JL, Chowdary P, Mutch NJ. Fibrinolytic abnormalities in acute respiratory distress syndrome (ARDS) and versatility of thrombolytic drugs to treat COVID-19. *J Thromb Haemost.* 2020;18(7):1548-1555.
44. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet.* 2020;395(10223):497-506.
45. Ghadge AA, Khaire AA. Leptin as a predictive marker for metabolic syndrome. *Cytokine.* 2019;121:154735.
46. Booth NA, Simpson AJ, Croll A, Bennett B, MacGregor IR. Plasminogen activator inhibitor (PAI-1) in plasma and platelets. *Br J Haematol.* 1988;70(3):327-333.
47. Morrow GB, Whyte CS, Mutch NJ. Functional plasminogen activator inhibitor 1 is retained on the activated platelet membrane following platelet activation. *Haematologica.* 2019;105:2824-2833.
48. Yatim N, Boussier J, Chocron R, et al. Platelet activation in critically ill COVID-19 patients. *Ann Intensive Care.* 2021;11(1):113.
49. Agrati C, Sacchi A, Tartaglia E, et al. The role of P-Selectin in COVID-19 coagulopathy: an updated review. *Int J Mol Sci.* 2021;22(15):7942.
50. Wu YP, Wei R, Liu ZH, et al. Analysis of thrombotic factors in severe acute respiratory syndrome (SARS) patients. *Thromb Haemost.* 2006;96(1):100-101.
51. Chen N, Zhou M, Dong X, et al. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study. *Lancet.* 2020;395(10223):507-513.
52. Gao Y, Li T, Han M, et al. Diagnostic utility of clinical laboratory data determinations for patients with the severe COVID-19. *J Med Virol.* 2020;92(7):791-796.
53. Mo P, Xing Y, Xiao Y, et al. Clinical characteristics of refractory COVID-19 pneumonia in Wuhan, China. *Clin Infect Dis.* 2020;73:e4208-e4213.
54. Chen G, Wu D, Guo W, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest.* 2020;130(5):2620-2629.
55. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet.* 2020;395(10229):1054-1062.
56. Prabhakaran P, Ware LB, White KE, Cross MT, Matthay MA, Olman MA. Elevated levels of plasminogen activator inhibitor-1 in pulmonary edema fluid are associated with mortality in acute lung injury. *Am J Physiol Lung Cell Mol Physiol.* 2003;285(1):L20-L28.
57. Ware LB, Matthay MA, Parsons PE, Thompson BT, Januzzi JL, Eisner MD. Pathogenetic and prognostic significance of altered coagulation and fibrinolysis in acute lung injury/acute respiratory distress syndrome. *Crit Care Med.* 2007;35(8):1821-1828.
58. Cheng M, Li Y, Wu J, et al. IL-8 induces imbalances between nitric oxide and endothelin-1, and also between plasminogen activator inhibitor-1 and tissue-type plasminogen activator in cultured endothelial cells. *Cytokine.* 2008;41(1):9-15.
59. He L, Ding Y, Zhang Q, et al. Expression of elevated levels of pro-inflammatory cytokines in SARS-CoV-infected ACE2+ cells in SARS patients: relation to the acute lung injury and pathogenesis of SARS. *J Pathol.* 2006;210(3):288-297.
60. Zhao X, Nicholls JM, Chen YG. Severe acute respiratory syndrome-associated coronavirus nucleocapsid protein interacts with Smad3 and modulates transforming growth factor-beta signaling. *J Biol Chem.* 2008;283(6):3272-3280.
61. Gralinski LE, Ferris MT, Aylor DL, et al. Genome wide identification of SARS-CoV susceptibility loci using the collaborative cross. *PLoS Genet.* 2015;11(10):e1005504.
62. Mestries JC, Kruthof EK, Gascon MP, Herodin F, Agay D, Ythier A. In vivo modulation of coagulation and fibrinolysis by recombinant glycosylated human interleukin-6 in baboons. *Eur Cytokine Netw.* 1994;5(3):275-281.
63. RECOVERY Collaborative Group. Tocilizumab in patients admitted to hospital with COVID-19 (RECOVERY): a randomised, controlled, open-label, platform trial. *Lancet.* 2021;397(10285):1637-1645.
64. Wise J. Covid-19: critically ill patients treated with arthritis drug tocilizumab show improved outcomes, researchers report. *BMJ.* 2020;371:m4530.
65. von Meijenfheldt FA, Havervall S, Adelmeijer J, Thalín C, Lisman T. Persistent endotheliopathy in the pathogenesis of long COVID syndrome: comment from von Meijenfheldt et al. *J Thromb Haemost.* 2022;20(1):267-269.

## SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

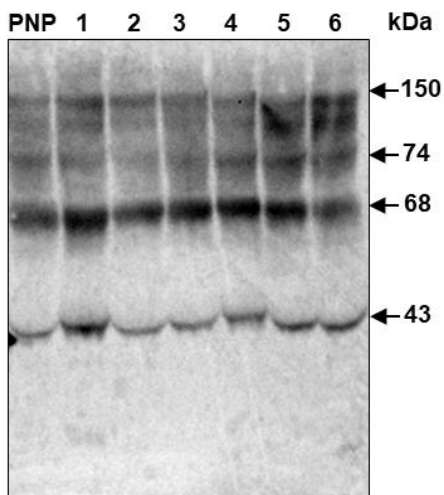
**How to cite this article:** Whyte CS, Simpson M, Morrow GB, et al. The suboptimal fibrinolytic response in COVID-19 is dictated by high PAI-1. *J Thromb Haemost.* 2022;20:2394-2406. doi: [10.1111/jth.15806](https://doi.org/10.1111/jth.15806)



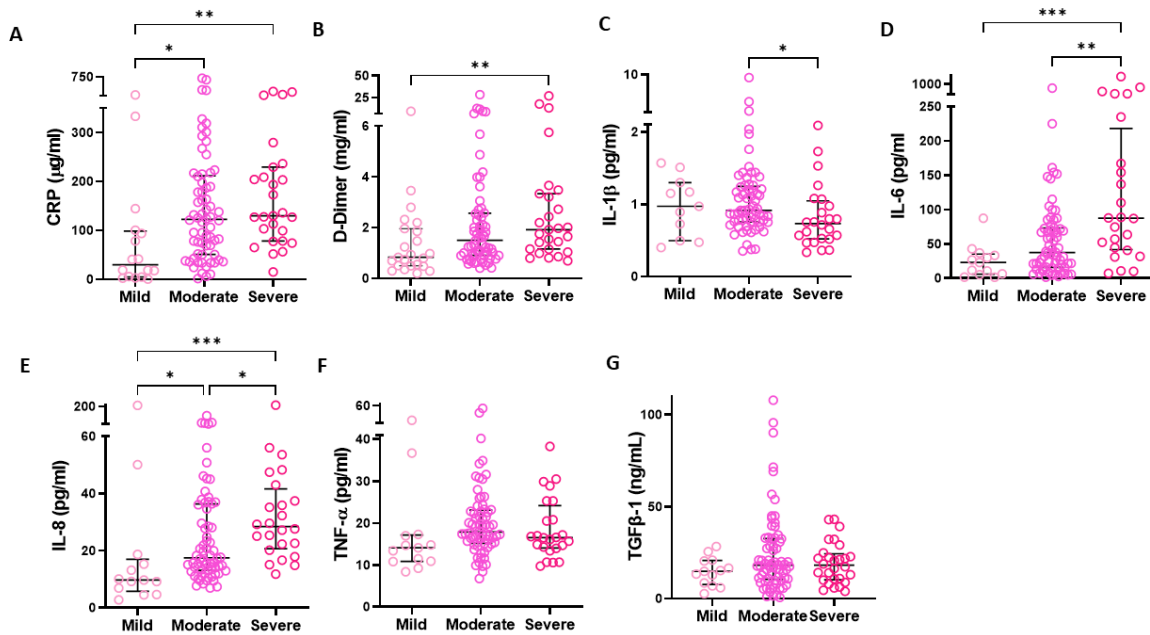
### Supplemental Material

Non-COVID-19 respiratory infection (24)	
LRTI/Pneumonia - Non-COVID-19	17
Viral Respiratory illness - Non-COVID-19	3
Aspiration Pneumonia	2
Exacerbation Asthma/COPD	2

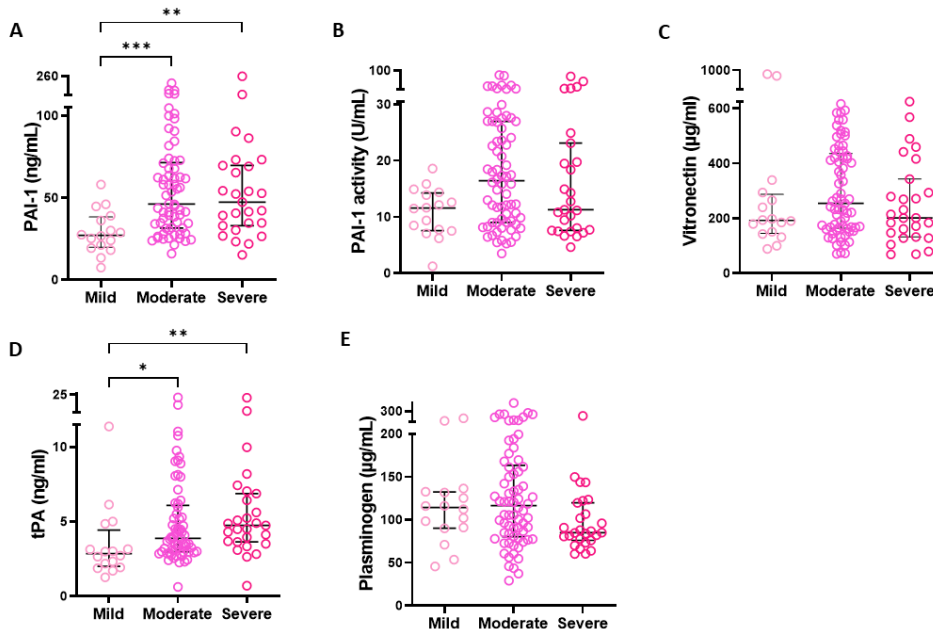
**Supplementary Table 1 – Breakdown of the type of Non-COVID-19 respiratory infections.** Lower respiratory tract infection (LRTI), chronic obstructive pulmonary disease (COPD).



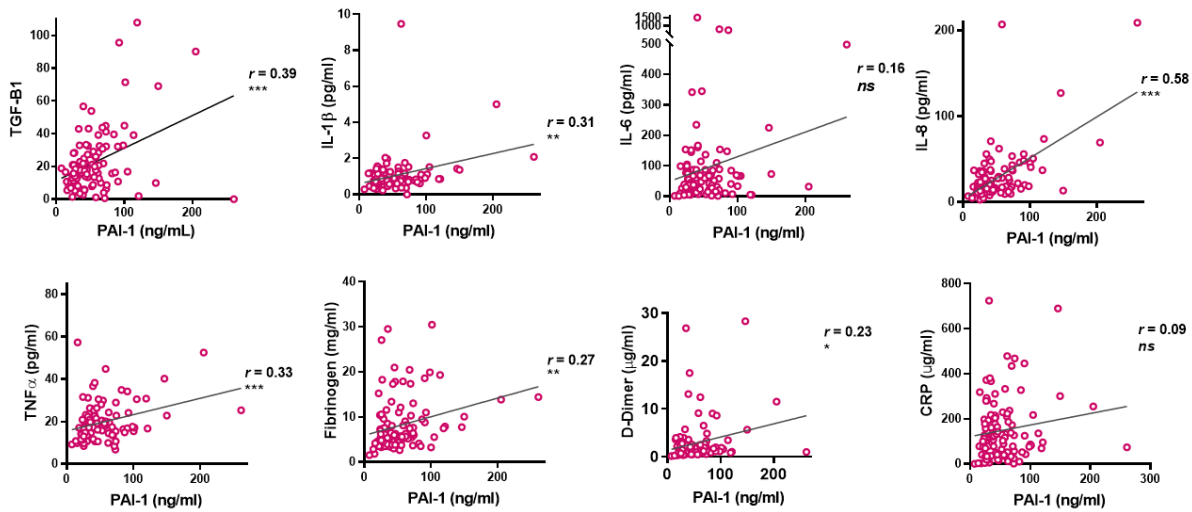
**Supplementary Figure 1 – PAI-1 – Vn complex detected by Western blotting.** COVID-19+ plasma samples (1-6) and pooled normal plasma (PNP) were separated on 4-12% NuPAGE Bis-Tris gels under non-reducing conditions, then transferred to PVDF membrane. PAI-1 was detected with polyclonal PAI-1 antibody (Affinity Biologicals) and donkey anti-sheep IgG conjugated to HRP (Sigma). Bands of PAI-1 protein were detected at 43, 68, 74 and 150 kDa.



**Supplementary Figure 2 – Inflammatory markers and proinflammatory cytokines increase with COVID-19 disease severity.** Inflammatory markers were grouped according into the following groups of disease progression; mild = patients that did not require supplemental oxygen, moderate = patients on supplemental oxygen support or severe = patients on high flow nasal cannula (HFNC), continuous positive airway pressure (CPAP) or patients that were ventilated. (A) CRP antigen, (B) D-dimer, (C) IL-1 $\beta$ , (D) IL-6, (E) IL-8, (F) TNF- $\alpha$  and (G) TGF $\beta$ -1. \*  $p < 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .

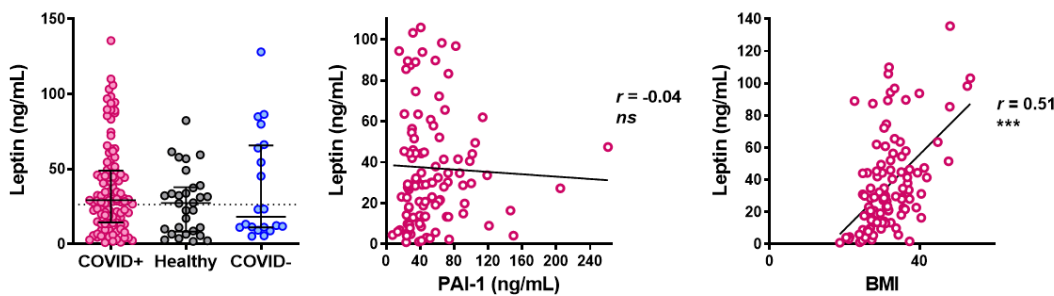


**Supplementary Figure 3 – PAI-1 and tPA increase with COVID-19 disease severity.** Fibrinolytic markers were grouped according to disease severity; mild = patients that did not require supplemental oxygen, moderate = patients on supplemental oxygen support or severe = patients on high flow nasal cannula (HFNC), continuous positive airway pressure (CPAP) or patients that were ventilated. (A) PAI-1 antigen, (B) PAI-1 activity, (C) vitronectin, (D) tPA antigen and (E) plasminogen. \*  $p < 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .



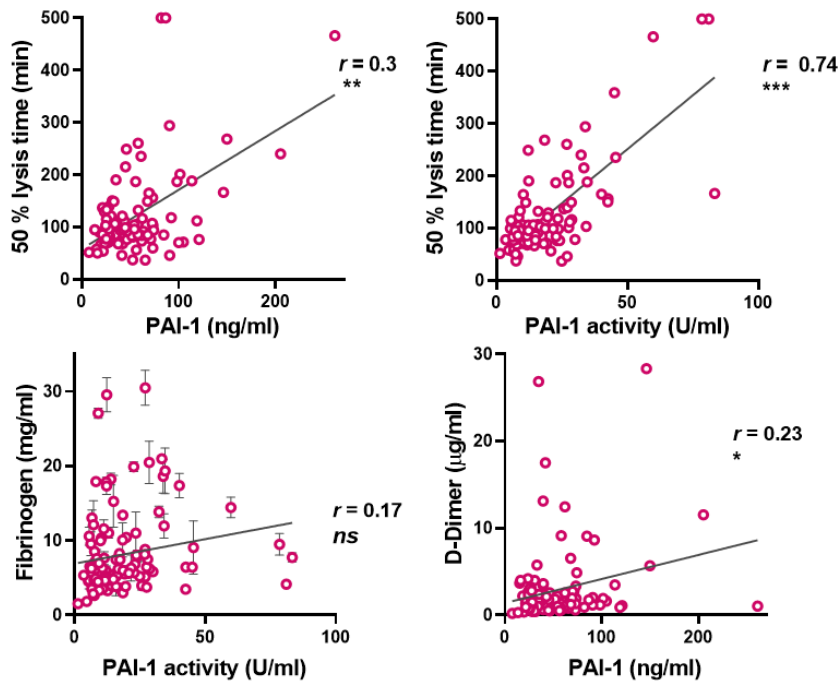
**Supplementary Figure 4 – PAI-1 correlates with inflammatory markers and proinflammatory cytokines.**

Pearson correlation coefficients ( $r$ ) were calculated between PAI-1 antigen and TGF- $\beta$ 1 - IL-1 $\beta$ , IL-6, IL-8 TNF- $\alpha$  fibrinogen, D-dimer and CRP. \*  $p < 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .



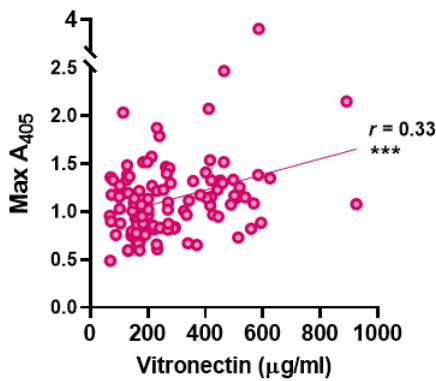
**Supplementary Figure 5 – Leptin plasma levels do not differ between COVID-19 positive and negative patients and does not correlate with PAI-1 levels.**

(A) The concentration of leptin (ng/mL) was measured in plasma from patients with COVID-19 (COVID+), other non-COVID-19 respiratory infections (COVID-) and healthy controls by Simple Plex™ assays on Ella™. Dotted lines indicate antigen concentrations in pooled normal plasma. Data are shown as median  $\pm$  interquartile range (IQR). Correlations of leptin with (B) PAI-1 and (C) BMI. \*\*\*  $p < 0.001$ , ns = non-significant.

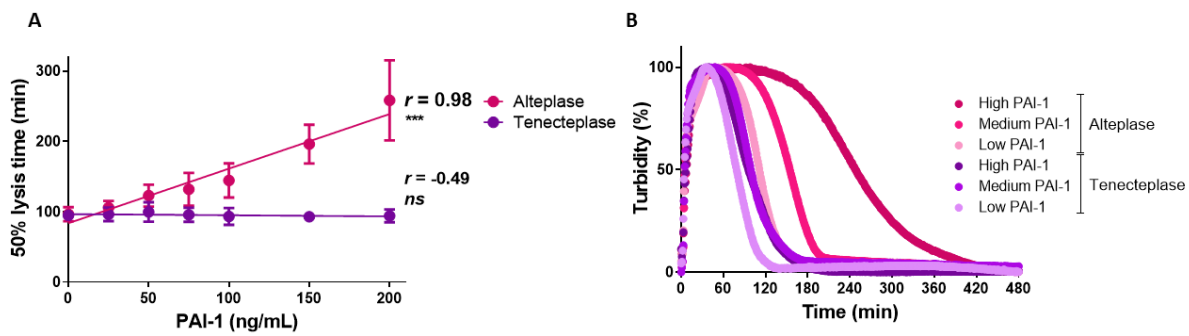


**Supplementary Figure 6 – PAI-1 correlates with fibrinolytic activity**

Pearson correlation coefficients ( $r$ ) were calculated between PAI-1 antigen or activity with tPA -mediated lysis and with fibrinogen and D-dimer and PAI-1 activity. \*  $p < 0.05$ , \*\*  $p < 0.01$  and \*\*\*  $p < 0.001$ .



**Supplementary Figure 7 – Vitronectin levels correlate with plasma clot maximum absorbance.** Pearson correlation coefficients ( $r$ ) were calculated between vitronectin and plasma clot maximum absorbance at 405nm. \*\*\*  $p < 0.001$ .



**Supplementary Figure 8 –PAI-1 delays clot lysis by Alteplase but not Tenecteplase.** Plasma clots (30 %) were formed in the presence of phospholipids (16 mM) ± Alteplase or Tenecteplase (300 pM). Clotting was initiated with CaCl<sub>2</sub> (10.6 mM) and thrombin (0.1 U/ml) and absorbance readings at 405 nm were taken every min for 8 h at 37 °C. (A) Purified PAI-1 was spiked into pooled plasma at 0, 25, 50, 75, 100, 150 and 200 ng/mL. Data show the median ± IQR time to 50 % lysis of plasma clots (n=4). The relationship between 50% lysis time and PAI-1 concentration was significantly different between the two plasminogen activators, Alteplase and Tenecteplase. \*\*\*  $p < 0.001$ . (B) Normalised clot lysis profiles of representative COVID-19 plasma with low <50 ng/mL [13.4-46.4 ng/mL]; medium 50-100 ng/mL [51.5 - 92.3 ng/mL] or high >100 ng/mL [103.3 - 260 ng/mL]) PAI-1 antigen (n=3 representative samples per PAI-1 grouping).