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1	The degradation of microcystin-LR using doped visible light		
2	absorbing photocatalysts.		
3			
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25 Abstract

Microcystins are one of the primary hepatotoxic cyanotoxins 26 released from cyanobacteria. The presence of these compounds in 27 water has resulted in the death of both humans and domestic and 28 29 wild animals. Although microcystins are chemically stable titanium 30 dioxide photocatalysis has proven to be an effective process for the removal of these compounds in water. One problem with this 31 process is that it requires UV light and therefore in order to develop 32 effective commercial reactor units that could be powered by solar 33 light it is necessary to utilize a photocatalyst that is active with 34 35 visible light. In this paper we report on the application of four visible light absorbing photocatalysts for the destruction of microcystin-LR 36 in water. The rhodium doped material proved to be the most 37 effective material followed by a carbon modified titania. The 38 39 commercially available materials were both relatively poor 40 photocatalysts under visible radiation while the platinum doped catalyst also displayed a limited activity for toxin destruction. 41

42

43 **1. Introduction**

Cyanobacterial toxins produced and released by cyanobacteria 44 in freshwater around the world are well documented (Carmichael, 45 1995; Sivonen, 1996). Microcystins are the most common of the 46 47 cyanobacterial toxins found in water, as well as being the ones most 48 often responsible for poisoning animals and humans who come into contact with toxic blooms and contaminated water (Codd et al., 49 1989; Dunn, 1996; Yu, 1994; Tsuji et al., 1995). The most 50 important toxin is microcystin-LR which is a cyclic heptapeptide 51 containing the amino acid 3-amino-9-methoxy-2,6,8-trimethyl-10-52 53 phenyldeca-4,6-dienoic acid (adda), with leucine and arginine in the Microcystins are chemically very stable and 54 variable positions. conventional water treatment processes have so far proved 55 relatively ineffective in removing them (Keijola et al., 1988, Lahti et 56 57 al., 1989, Lawton and Robertson, 1999). The formation of 58 chlorinated by-products of the microcystins has been reported by Merel et al. (2009). This group reported a detailed study of the use 59 of chlorination as a treatment process for the removal of 60 microcystins from water. Chlorination, this is only effective at 61 62 relatively high doses with a sufficient contact time. Although chlorinated by-products have been characterised as yet the toxicity 63 biodegradation of by-products is unknown. The 64 of these microcystins -LR, -LF and the pentapeptide toxin nodularin in water 65 66 has previously been studied (Edwards et al., 2008; Torunska et al.,

67 2008) who reported degradation half lives for the toxins of between 68 4 and 18 d. Moreover the use of methods such as granular carbon 69 filtration and photochemical degradation have shown only limited 70 efficacy (Kaya and Sano, 1998, Reynolds, 1987; Tsuji et al., 1994). 71 Consequently there is a growing need for effective treatment 72 technologies for removal of microcystins from water.

73

We previously reported the effectiveness of TiO_2 74 have photocatalysis for the removal of microcystins in water (Robertson 75 et al., 1997; Robertson et al., 1998; Lawton and Robertson, 1999; 76 77 Robertson et al., 1999; Lawton et al., 1999; Cornish et al., 2000, Liu et al., 2002; Liu et al., 2003). Although the process has been 78 demonstrated to be extremely effective in removing the toxin a 79 limitation of the process was the fact that the catalyst requires UV 80 81 light. If a suitable visible light absorbing photocatalyst could be prepared the viability of the photocatalytic process would be 82 significantly improved as there could even be the potential of 83 operating the process using sun light. Choi et al. (2007) reported 84 the use of a nitrogen doped TiO₂ catalyst for the destruction of 85 86 microcystin under visible irradiation. Using this catalyst approximately 50% of a 5 μ M solution of microcystin-LR was 87 degraded in 30 min with virtually complete destruction being 88 achieved in 2 h. Pelaez et al. (2009) studied the photocatalytic 89 destruction of microcystin-LR using visible light absorbing N-F-90

91 codoped TiO₂ nanoparticles. At pH 3 the toxin was completed degraded within 300 min when irradiated with light at 420 nm. It 92 was proposed that under these conditions the electrostatic 93 interaction between the toxin and photocatalyst favoured the 94 decomposition process. The purpose of the research detailed in this 95 96 paper was to look at the potential effectiveness of a number of visible absorbing photocatalysts for the destruction of microcystin-97 LR. The efficiency of these materials was also compared to the 98 material that has been the subject of our previous work on 99 photocatalytic destruction of microcystin, the UV light absorbing 100 101 Degussa P25 TiO₂.

102

103 2. Materials and experimental methods

104 2.1. Materials

105 The two unmodified titania samples P25 (Degussa) and TH (Titanhydrat, Kerr-McGee) were commercially available, whereas 106 the modified samples TiO_2 -C (TiO_2 -C1b: Sakthivel and Kisch, 2003), 107 ${[Ti]OPtCl_4L}^{n-}$, L = H₂O, OH⁻, n = 1, 2, (abbreviated as TiO₂-108 Pt(IV)) (Burgeth and Kisch, 2002), and ${[TiO_2]-O-RhCl_3(H_2O)_2}^-$ 109 110 (abbreviated as TiO₂-Rh(III)) (5% RhCl₃/TH: Dai et al., 2008) were prepared according to the literature. P25 TiO₂ was used as a 111 standard material for comparison on the relative effectiveness of 112 the catalysts detailed above. This has previously been established 113 114 as the most effective UV photocatalyst for the destruction of 115 cyanotoxins (Liu et al., 2009). Microcystin-LR was prepared as 116 previously reported (Edwards et al., 1996). The toxin was 117 resuspended in water prior to use.

118

119 2.2 Photocatalysis

The photocatalysis experiments were performed in 20 mL glass 120 vials under constant stirring at a distance of 30 cm from the front of 121 the light source (500 W Halogen, irradiation 393 μ M⁻¹, Temp 306 K 122), as illustrated in Fig. 1. A 2% w/v sodium nitrite filter was used to 123 eliminate and UV photons generated by the halogen source entering 124 125 the reaction vessel. An IR filter was also used to minimise any thermal effects on the process. 50 mg of the photocatalysts were 126 added to 5 mL of a 0.1 mg mL⁻¹ microcystin-LR solution. The 127 photocatalysis was conducted in Milli-Q water (18.2 m Ω cm) and 128 129 the reaction pH was 4. After mixing for 5 min an initial sample was 130 removed to determine extent of dark adsorption. From previous work it has been established that the dark adsorption process of 131 microcystins on titania photocatalysts is rapid and equilibrium is 132 established within the first five min (Robertson et al., 1997; Lawton 133 et al., 2003). The photocatalysis was then initiated and samples 134 were removed at 10 min intervals for the first 60 min after which 135 sampling was performed every 20 min to a maximum photocatalysis 136 period of 120 min. 137

138

139 *2.3* Analysis

140 Treated samples were analysed by HPLC with photodiode array
141 detection as previously detailed (Lawton et al., 1994).

142

143 **3. Results and discussion**

144 In order to ensure that the visible activity of the photocatalysts was being observed and there was no contribution to the process 145 from UV light, the process was initially examined using a standard 146 Degussa P25 material. On irradiating the suspension of P25 TiO₂ 147 and microcystin with the halogen light filtered through 2% sodium 148 149 nitrite filter, no significant level of toxin degradation was observed up to 120 min photocatalysis (Fig. 2a). The results of this initial 150 study confirmed that all residual UV light generated by the tungsten 151 halogen source had been removed, so only visible photons were 152 153 entering the reactor cell. Prior to using this filter set up a modest 154 destruction of microcystin-LR was achieved with unmodified photocatalyst materials from the residual UV in the halogen source. 155 In order to determine the true level of visible activity of the 156 modified photocatalyst material, it was therefore necessary to utilise 157 158 this effective UV filter in order to eliminate any potential UV 159 activation of the materials.

160

161 The photocatalytic activity of the five catalysts for the 162 destruction of microcystin-LR was subsequently assessed in the

163 same experimental setup. On adding the photocatalyst to the microcystin solution it was initially clear that each of the materials 164 displayed varying levels of initial dark adsorption of the toxin. This 165 dark adsorption of the toxins on TiO₂ photocatalysts has been 166 167 previously reported and is believed to be due to a combination of 168 the surface charge of the photocatalysts material and the hydrophobicity of the toxin (Lawton et al., 2003). Table 1 169 summarises for each of the photocatalysts the amount of 170 microcystin remaining in solution after dark adsorption and after 60 171 min of irradiation time. The photocatalytic destruction of the 172 173 microcystin over 120 min irradiation time for both modified and unmodified materials under visible light irradiation is detailed in Fig. 174 2b. At the high photocatalyst concentration employed (10 g L^{-1}) the 175 amount of absorbed light should be about the same in each 176 177 experiment and reaction rates are therefore comparable. Unlike the two unmodified titania samples, which do not absorb visible light, all 178 modified photocatalysts efficiently decomposed the toxin, with TiO₂-179 C and TiO₂-Rh(III) having the highest activity. In the latter system 180 half of the toxin present in solution was degraded after 10 min 181 182 whereas TiO₂-C needed 30 min to induce the same amount of destruction. As can be recognized, only the TiO₂-Rh(III) sample 183 enabled complete toxin degradation after 60 min. 184

185

186 Scheme 1 summarizes the positions of band edge and absorption onset of the various photocatalysts. It is rather unlikely 187 that the small changes in the redox potentials of the light-generated 188 electron and holes are responsible for the different degradation 189 activities. It is more likely that the different chemical nature of the 190 191 reactive holes seems to be decisive. It was proposed that in the case of the Rh(III) modified material the hole is a Rh(IV) species 192 (Eq. 1, Dai et al., 2008), whereas in TiO₂-C and related carbon-193 modified titania it may be a radical or radical cation of a condensed 194 aromatic compound (Neumann et al., 2005; Zabek et al., 2009 and 195 196 references cited therein). It appears very probable that such different species oxidize microcystin with different reaction rates. It 197 is known that the interfacial electron transfer from the conduction 198 band to oxygen (Eq. 2) is the rate-determining step of most 199 200 semiconductor catalyzed photooxidation reactions (Gerischer and Heller, 1991; Schwitzgebel et al., 1995). If this would be the case, 201 then TiO₂-C should react faster than TiO₂-Rh(III) since the driving 202 force is lower for the latter due to the corresponding flatband 203 positions of -0.48 V (Sakthivel and Kisch, 2003) and -0.34 V (Dai 204 205 et al., 2008) for the carbon- and rhodium-modified material, 206 respectively. However, it cannot be excluded that charge generation in TiO₂-Rh(III) is more efficient resulting in a higher 207 electron concentration and therefore in faster oxygen reduction (Eq. 208 209 2). As a result of this also the concentration of the hydroxyl radical,

the other strongly oxidizing intermediate generated through reaction steps according to Eq. 3-6, are expected to be higher than in TiO₂-C.

213

214
$$[TiO_2]O-Rh^{3+} + hv \rightarrow [TiO_2]O-Rh^{4+} + e_{CB}$$
 (1)

215
$$O_2 + e_{CB} \rightarrow O_2^{-\bullet}$$
 (2)

216
$$O_2^{-\bullet} + H^+ \rightarrow HO_2^{\bullet}$$
 (3)

217
$$HO_2^{\bullet} + HO_2^{\bullet} \rightarrow H_2O_2 + O_2$$
 (4)

218
$$H_2O_2 + O_2^{-\bullet} \rightarrow OH^{\bullet} + OH^{-} + O_2$$
 (5)

219
$$H_2O_2 + e_{CB} \rightarrow OH^{\bullet} + OH^{-}$$
 (6)

220

221 The observation that TiO_2 -Pt(IV) is much less active than the 222 rhodium and carbon modified samples suggests that the adsorbed 223 chlorine radical (representing the reactive hole) generated through a homolytic Pt-Cl bond cleavage (Burgeth and Kisch, 2002) can 224 225 induce only an inefficient microcystin oxidation. TiO₂-Pt(IV) exhibits the strongest dark adsorption of 84%. A light-induced desorption of 226 227 microcystin may be responsible for the small concentration increase found for P25. 228

229

In the work performed by Choi et al. (2007) using a nitrogen doped titania sample 50% toxin destruction was achieved in around 30 min with complete decomposition reported within 2 h visible light photocatalysis. It is not possible to directly compare the results of

this study with those obtained in the work of Choi et al. (2007) as
the experimental conditions such as the initial toxin concentration
and light intensity were not the same.

237

In order to assess the relative efficiency of the visible light 238 photocatalytic process to the UV absorbing titania, the degradation 239 of microcystin-LR was examined using both a standard Degussa P25 240 material and a modified P25 catalyst TiO₂-Pt(IV) under UV 241 irradiation. For both materials the microcystin was decomposed 242 243 within 10 min of photocatalysis (Fig. 2c) as compared to about 20 min for the best visible light photocatalyst TiO₂-Rh(III). This 244 unexpected result can be rationalized by assuming that holes 245 generated below the valence band edge ("hot carriers") rather 246 oxidize microcystin than relax to the conduction band edge level. 247 The opposite behaviour may be responsible for the lower activity of 248 249 TiO_2 -Rh(III).

250

The use of artificial UV light sources for photocatalytic water treatment represents one of the most significant costs of the process. Consequently there has been significant interest in utilizing sunlight as the energy source for large scale photocatalytic water treatment (Bahnemann, 2004). Unfortunately UV light represents approximately 5% of the energy in solar light, while 45% is in the visible region. Therefore at the planet's surface, the typical UV-flux

of 20 to 30 W m⁻², the sun generates 0.2 to 0.3 mol photons m⁻² h⁻¹ 258 259 between 300-400 nm for potential utilisation by photocatalytic reactors (Bahnemann, 1994). If visible light absorbing catalysts 260 were to be utilised potentially up to nine times this energy could be 261 available, depending on the band gap of the catalyst being utilized. 262 263 Consequently, although it would appear in this study that the TiO_2/UV process is more efficient than the TiO_2/V is process, the 264 advantages of being able to utilise visible light far outweigh the 265 relatively poorer photocatalytic efficiency particularly for any large 266 scale application. There may be potential toxicity risks associated 267 268 with using metal doped titania materials. This risk would depend on the potential leaching of the dopant metals from the titania material 269 into the water. In any commercial application it would be necessary 270 to assess the risk of such a leaching process occurring and this 271 272 would need to be a subject of more in depth future work.

273

4. Conclusions

The photocatalytic degradation of the cyanotoxin, microcystin-LR has been achieved using visible light absorbing photocatalysts. The process was demonstrated to be particularly efficient for the TiO₂-Rh(III) material with 90% of the toxin being destroyed in 20 min irradiation. Although the TiO₂/UV process was still relatively more efficient, the advantages of being able to use visible light to drive this process significantly outweigh the marginally faster

kinetics under UV irradiation. These visible light absorbing materials
could therefore significantly improve the viability of semiconductor
photocatalysis as a practical water treatment process for removal of
microcystins from potable water supplies.

286 **References**

- 287 Bahnemann, D.W., 1994. Solare Abwasserentgiftung. Nachr.
- 288 Chem. Tech. Lab. 42, 378-388.
- Bahnemann, D.W., 2004. Photocatalytic water treatment: Solar
 energy applications. Sol. Energy 77, 445-459.
- Burgeth, G., Kisch, H., 2002. Photocatalytic and
- 292 photoelectrochemcial properties of titania-chloroplatinate(IV).
 293 Coord. Chem. Rev. 230, 41-47.
- 294 Carmichael, W.W., 1995. Cyanobacterial toxins. In: Hallegraeff,
- G.M., Anderson, D.M., Cembella, A.D. (Eds.). Manual on
 Harmful Marine Microalgae. UNESCO, Paris, pp. 163-175.
- 297 Choi, H., Antoniou, M.G., Pelaez, M., De la Cruz, A.A., Shoemaker,
- J.A., Dionysiou, D.D., 2007. Mesoporous nitrogen-doped
 TiO₂ for the photocatalytic destruction of the cyanobacterial
 toxin microcystin-LR under visible light irradiation. Environ.
 Sci. Technol. 41, 7530-7535.
- Codd, G.A., Bell, S.G., Brooks, W.P., 1989. Cyanobacterial toxins in
 water. Water Sci. Technol. 21(3), 1-13.
- Cornish, B.J.P.A., Lawton, L.A., Robertson, P.K.J., 2000. Hydrogen
 peroxide enhanced photocatalytic destruction of microcystin-
- 306 LR. Appl. Catal. B-Environ. 25, 59-67.
- 307 Dai, Zh., Burgeth, G., Parrino, F., Kisch, H., 2008. Visible light
- 308 photocatalysis by a Titania–Rhodium(III) complex. J.
- 309 Organometal. Chem. 694, 1049-1054.

- 310 Dunn, J., 1996. Algae kills dialysis patients in Brazil. Brit. Med. J.
- 311 **312, 1183-1184**.
- 312 Edwards, C., Lawton, L.A., Coyle, S.M., Ross, P., 1996. Laboratory-
- 313 scale purification of microcystins using flash chromatography
- and reversed-phase high-performance liquid chromatography.
- 315 J. Chromatogr. A 734, 163-167.
- Edwards, C., Graham, D., Fowler, N., Lawton, L.A., 2008.
- Biodegradation of microcystins and nodularin in freshwaters.
- 318 Chemosphere 73, 1315-1321.
- 319 Gerischer, H., Heller, A., 1991. The role of oxygen in photooxidation
- 320 of organic molecules on semiconductor particles. J. Phys.
- 321 Chem.-US 95, 5261-5267.
- 322 Kaya, K., Sano, T., 1998. A photodetoxification mechanism of the
- 323 cyanobacterial hepatotoxin microcystin-LR by ultraviolet

324 irradiation. Chem. Res. Toxicol. 11, 159-163.

- 325 Keijola, A.M., Himberg, K., Esala, A.L., Sivonen, K., Hiisvirta, L.,
- 326 1988. Removal of cyanobacterial toxins in water treatment
- 327 processes: laboratory and pilot-scale experiments. Toxic.
- 328 Assess. 3, 643-656.
- 329 Lahti, K., Hiisvirta, L., 1989. Removal of cyanobacterial toxins in
- 330 water treatment processes: Review of studies conducted in
- Finland. Water Supply Manage. 7, 149-154.
- Lawton, L.A., Edwards, C. Codd, G.A., 1994. Extraction and high
- 333 performance liquid chromatographic method for determination

- of microcystins in raw and treated waters. Analyst 119, 1525–
 1530.
- Lawton, L.A., Robertson, P.K.J., 1999. Physico-chemical methods
 for the removal of microcystins. Chem. Soc. Rev. 28, 217224.
- 339 Lawton, L.A., Robertson, P.K.J., Cornish, B.J.P.A., Jaspars, M.,
- 340 1999. Detoxification of microcystins (cyanobacterial
- 341 hepatotoxins) using TiO₂ photocatalytic oxidation. Environ.
- 342 Sci. Technol. 33, 771-775.
- 343 Lawton, L.A., Robertson, P.K.J., Cornish, B.J.P.A., Marr, I.L.,
- Jaspars, M., 2003. Processes influencing surface interaction
- and photocatalytic destruction of microcystins on titanium
 dioxide photocatalysts. J. Catal. 213, 109-113.
- Liu, I., Lawton, L.A., Cornish, B.J.P.A., Robertson, P.K.J., 2002.
- 348 Mechanistic and toxicity studies of the photocatalytic oxidation 349 of microcystin-LR. J. Photoc. Photobio. A 148, 349-354
- Liu, I., Lawton, L.A., Robertson, P.K.J., 2003. Mechanistic studies of

351 the photocatalytic oxidation of microcystin-LR: An

- 352 investigation of by-products of the decomposition process.
- 353 Environ. Sci. Technol. 37, 3214-3219.
- Liu, I., Lawton, L.A., Bahnemann, D.W., Proft, B., Robertson, P.K.J.,
- 355 2009. The photocatalytic decomposition of microcystin-LR
- using selected titanium dioxide materials. Chemosphere 76,
- **549-553**.

359	Merel, S., LeBot, B., Clément, M., Seux, R., Thomas, O., 2009. MS
360	identification of microcystin-LR chlorination by-products.
361	Chemosphere 74, 832-839.
362	Neumann, B., Bogdanoff, P., Tributsch, H., Sakthivel, S., Kisch, H.,
363	2005. Electrochemical mass spectroscopic and surface
364	photovoltage studies of catalytic water photooxidation by
365	undoped and carbon doped titania. J. Phys. Chem. B 109,
366	16579-16586.
367	Pelaez, M., de la Cruz, A.A., Stathatos, E., Falaras, P., Dionysiou,
368	D.D., 2009. Visible light-activated N-F-codoped TiO_2
369	nanoparticles for the photocatalytic degradation of
370	microcystin-LR in water. Catal. Today 144, 19-25.
371	Reynolds, C.S., 1987. Cyanobacterial water blooms. In: Callow, J.A.
372	(Ed.). Advances in Botanical Research. Academic Press,
373	London, vol. 13, pp. 67-143.
374	Robertson, P.K.J., Lawton, L.A., Munch, B., Rouzade J., 1997.
375	Destruction of cyanobacterial toxins by semiconductor
376	photocatalysis. Chem. Commun. 393-394.
377	Robertson, P.K.J., Lawton, L.A., Cornish, B.J.P A., Jaspars, M.,
378	1998. Processes influencing the destruction of microcystin-LR
379	by TiO ₂ photocatalysis. J. Photoc. Photobio. A 116, 215-219.

380	Robertson, P.K.J., Lawton, L.A., Munch, B., Cornish, B.J.P.A., 1999.
381	The destruction of cyanobacterial toxins by TiO_2
382	photocatalysis. J. Adv. Oxid. Technol. 4, 20-26.
383	Sakthivel, S., Kisch, H., 2003. Daylight photocatalysis by carbon-
384	modified titanium dioxide. Angew. Chem. Int. Edit. 42, 4908-
385	4911.
386	Schwitzgebel, J., Ekerdt, J. G., Gerischer, H., Heller, A., 1995. Role
387	of the oxygen molecule and of the photogenerated electron in
388	TiO2-photocatalyzed air oxidation reactions. J. Phys. Chem
389	US 99, 5633-5638.
390	Sivonen, K., 1996. Cyanobacterial toxins and toxin production.
391	Phycologia 35, 12-24.
392	Torunska, A., Bolalek, J., Plinski, M., Mazur-Marzec, H., 2008.
393	Biodegradation and sorption of nodularin (NOD) in fine-
394	grained sediments. Chemosphere 70, 2039-2046.
395	Tsuji, K., Nalto, S., Kondo, F., Ishikawa, N., Watanabe, M.F., Suzuki
396	M., Harada, KI., 1994. Stability of microcystins from
397	cyanobacteria: Effect of light on decomposition and
398	isomerization. Environ. Sci. Technol. 28, 173-177.
399	Tsuji, K., Watanuki, T., Kondo, F., Wanatabe, M., Suzuki, S.,
400	Nakazawa, H., Suzuki, M., Uchida, H., Harada, K., 1995.
401	Stability of microcystins from cyanobacteriaII. effect of UV
402	light on decomposition and isomerization. Toxicon 33, 1619-
403	1631.

Yu, S.-Z., 1994. Blue-green algae and liver cancer. In: Steffensen,
D.A, Nicholson, B.C. (Eds.). Toxic Cyanobacteria: Current
Status of Research and Management. Australia Centre for
Water Quality Research, Salisbury, Australia, pp. 75-85.
Ząbek, P., Eberl, J., Kisch, H., 2009. On the origin of visible light
activity in carbon-modified titania. Photoch. Photobio. Sci. 8,
264–269.

412 List of Captions for Figures.

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Fig. 1. Experimental set up for Photocatalysis Studies.

Fig. 2. a) Degradation of microcystin-LR in the presence of P25 with no illumination. b) Degradation of microcystin-LR using different photocatalyst materials. TH \blacktriangle , P25 \blacksquare , TiO₂-C \blacklozenge , TiO₂-Pt(IV) \square and TiO₂-Rh(III) \diamondsuit . c) Degradation of microcystin-LR in the presence of modified P25 TiO₂-Pt(IV) \blacksquare and P25 \blacklozenge under UV illumination using a deuterium light source.

423 **Scheme 1**. Band edge positions and onset of absorption (---) 424 of various photocatalysts. Lengths of arrows correspond to energy 425 of exciting light ($\lambda \ge 400$ nm). (Sakthivel and Kisch, 2003; Burgeth 426 and Kisch, 2002; Dai et al., 2008.)

427



Table 1. Percentage of Microcystin-LR in solution afterequilibration in the dark (a) and after 60 min of visible light

irradiation (b).

Photocatalyst	% Microcystin-LR remaining in		
	solution		
	t = 0	T = 60 min	
P25	57	60	
ТН	80	70	
TiO ₂ -Pt(IV)	16	10	
TiO ₂ -C	68	20	
TiO ₂ -Rh(III)	40	0	





Scheme 1.