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New directions and challenges in engineering biologically-enhanced biochar for biological water treatment.

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1 **New directions and challenges in engineering Biologically-Enhanced**
2 **Biochar for biological water treatment**

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26 **Abstract**

27 Cost-effective, efficient, and sustainable water treatment solutions utilising existing materials
28 and technology will make it easier for low and middle-income countries to adopt them,
29 improving public health. The ability of biochar to mediate and support microbial degradation
30 of contaminants, combined with its carbon-sequestration potential, has attracted attention in
31 recent years. Biochar is a possible candidate for use in cost-effective and sustainable
32 biological water treatment, especially in agrarian economies with easy access to abundant
33 biomass in the form of crop residues and organic wastes. This review evaluates the scope,
34 potential benefits (economic and environmental) and challenges of sustainable biological
35 water treatment using ‘Biologically-Enhanced Biochar’ or BEB. We discuss the various
36 processes occurring in BEB systems and demonstrate the urgent need to investigate microbial
37 degradation mechanisms. We highlight the need to correlate biochar properties to biofilm
38 development, which can eventually determine process efficiency. We also demonstrate the
39 various opportunities in adopting BEB as a cheaper and more viable alternative in Low and
40 Middle Income Countries and compare it to the current benchmark, ‘Biological Activated
41 Carbon’. We focus on the recent advances in the areas of data science, mathematical
42 modelling and molecular biology to systematically and sustainably design BEB filters, unlike
43 the largely empirical design approaches seen in water treatment. ‘Sequential biochar systems’
44 are introduced as specially designed end-of-life techniques to lower the environmental impact
45 of BEB filters and examples of their integration into biological water treatment that can fulfil
46 zero waste criteria for BEBs are given.

47 **Keywords:** Biologically-Enhanced Biochar, Biological Water Treatment, Biochar-Microbial
48 interactions, Sequential Biochar Systems

49

50 **1. Introduction**

51

52 According to the World Health Organisation, half of the world's population will be living in
53 water-stressed areas by 2025¹. Only 8% of the wastewater in low-income countries is
54 currently being treated and at least 2 billion people use a drinking water source contaminated
55 with faeces leading to 485 000 deaths from diarrhoea each year¹⁻³. With climate crisis
56 looming in the distance, there is an urgent need for a massive collaborative effort rooted in
57 inter-disciplinary research to find affordable and sustainable solutions for wastewater
58 treatment. It is in this regard that we review the scope of using specially designed
59 'Biologically-Enhanced Biochar' or BEB in sustainable biological water treatment.

60 Biochar is a carbon-rich product resulting from thermochemical conversion of carbonaceous
61 materials in an oxygen-deficient environment at high temperatures and is a carbon-negative
62 technology (300-800°C) ⁴⁻⁷. Recent developments in biochar research highlight the potential
63 for many applications ranging from water treatment, soil remediation, and agriculture to
64 energy conversion and storage ^{4,8-10}. There is increasing evidence of biochar mediation in
65 microbial metabolism in several energy and environmental applications such as soil and
66 water remediation, anaerobic digestion, and several Microbial Electrochemical Technologies
67 (METs) such as Microbial Fuel Cells (MFCs) ¹¹⁻¹⁴. Biochar mediation in microbial
68 degradation of contaminants combined with adsorbent properties make biochar a very
69 attractive option for biological water treatment.

70 Carbonaceous materials such as biochar can be used in tertiary water treatment which use
71 both adsorption and microbial activity to remove contaminants ^{5,15,16}. Here, microbes
72 (naturally present in water or externally introduced to suit target contaminants) can
73 immobilise on the surface of porous carbon forming biofilms. Biofilms then metabolise and
74 degrade the adsorbed contaminants in a bio-regeneration process. This bioregeneration

75 process frees up clogged pores of the filter material (such as biochar), regenerating their
76 adsorptive capacity and significantly increasing their lifespan^{17,18}. Despite limited experience
77 with the BEBs in biological water treatment, this review discusses the potential and
78 opportunities to deploy biochar for efficient, cost-effective and sustainable water treatment
79 applications, especially for on-site and home-scale water treatment units in Low and Middle-
80 Income Countries (LMICs) where centralised, industrial-scale water treatment units may be
81 too expensive to operate.

82 Moreover, current water treatment units are largely designed based on empirical research and
83 there is no clear consensus on how, why and to what extent biochar-microbial interactions
84 can affect the efficiency of the water treatment application. Recent technological
85 advancements and a wide array of analytical, statistical, mathematical, and molecular biology
86 techniques allow us to decode biochar-microbial interactions more systematically, making
87 this review timely and important. This review examines the dynamic and complex biofilm-
88 biochar interactions of BEB in water treatment and identifies gaps that require new research.
89 The review provides directions that will allow researchers to navigate through this highly
90 multi-disciplinary area and help bring affordable biochar-based water treatment solutions to
91 everyone, especially to people in LMICs. We discuss opportunities for adopting BEBs and
92 compare biochar with activated carbon (a benchmark material in tertiary water treatment) in
93 economic and environmental aspects. Using the multi-functionality of biochar, we
94 demonstrate how ‘sequential biochar systems’ can be integrated in biological water treatment
95 systems to meet zero waste criteria¹⁹. In this way, BEB filters have the potential to be safely
96 and effectively used in sequence or combination with various other applications such as
97 MFCs, anaerobic digestion, soil application and gas adsorption.

98 **2. BEB processes for tertiary water treatment integrating adsorption and** 99 **biodegradation**

100 Most industrial and municipal water treatment processes (drinking water treatment units,
101 waste water treatment units) comprise of primary, secondary and tertiary treatments to
102 remove organic, inorganic and biological contaminants ^{5,15}. Primary treatments mostly use
103 physical process such as coagulation and flocculation (with aluminium, ferric salts) and
104 chemical precipitation to remove solids, while secondary treatments generally involve
105 chemical and biological processes such as aerobic and anaerobic reactors to remove organic
106 matter ^{20,21}. Biological Activated Carbon (BAC) is commonly used as part of advanced water
107 treatments for combined adsorption and biodegradation of predominantly organic matter, but
108 also other inorganic and biological pollutants which could not be removed in primary and
109 secondary treatments⁵. In a typical BAC process, activated carbon acts as a support for
110 artificially introduced or naturally occurring microorganisms. These immobilised microbes
111 can reproduce on the activated carbon surface eventually forming BAC and facilitate
112 contaminant removal via a combined adsorption and biodegradation process ^{5, 17}. Other
113 tertiary treatments such as ion-exchange resins, photocatalysis, membrane processes,
114 disinfection (chlorination, ozonation) are also used ^{17,21,22}. Ozonation and chlorination steps
115 may precede or succeed BAC treatment depending on the process requirements ²³.

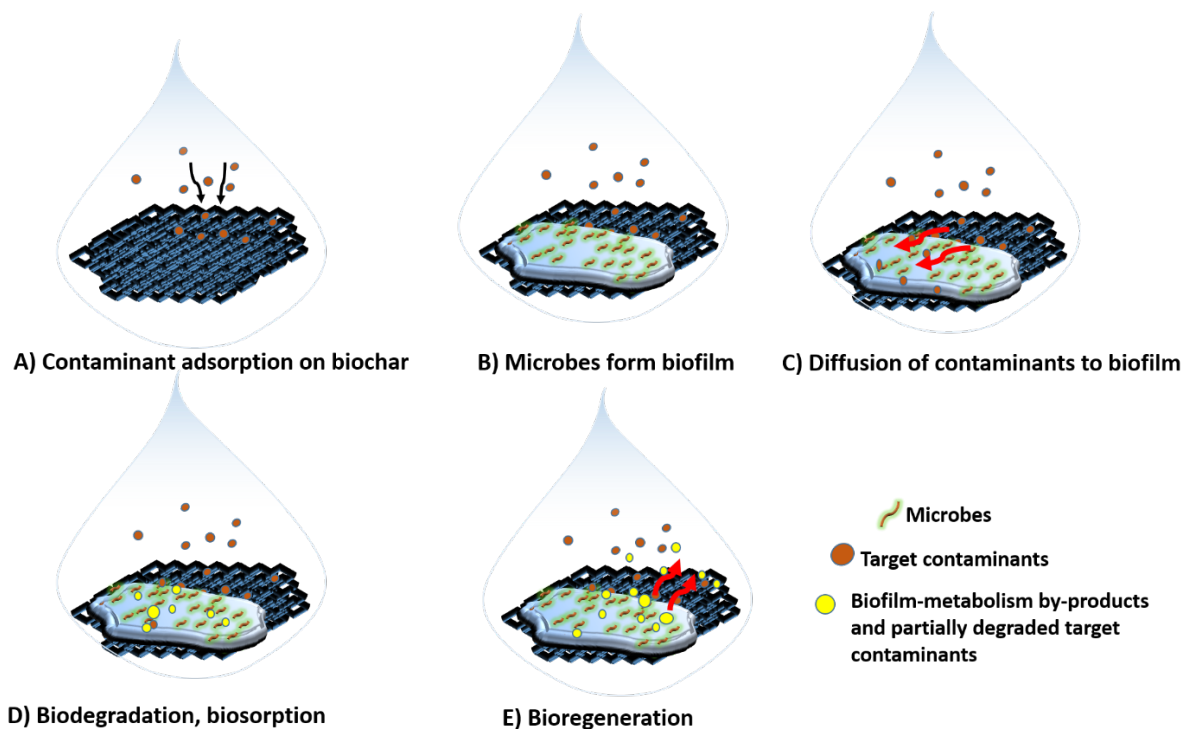
116 Biological water treatment using BEB, like BAC processes, can be used as tertiary water
117 treatment in sequence with other water treatment steps or in stand-alone on-site water
118 treatment units. The biofilms can degrade and remove a wide range of organic, inorganic and
119 biological water-borne contaminants ^{18,24,25}. This combined adsorption and bio-regeneration
120 process has shown to reduce the BAC costs by a factor of 2-3 in comparison with the use of
121 AC for adsorption-based removal alone ²⁶. Çeçen et.al. comprehensively detailed the history,

122 mechanisms and mathematical models of this integrated approach, both for attached-growth
123 (BAC) and suspended-growth in Powdered Activated Carbon (PAC) treatment ⁵. Here we
124 focus on attached-growth biofilm systems on biochar for water treatment, since powdered
125 carbon processes are more energy intensive owing to the extra step needed to separate the
126 suspended powders from the effluent after water treatment and are therefore less relevant in
127 LMICs ⁵.

128 **3. Biological water treatment using BEB systems**

129 **3.1 Adsorption, Biosorption and Biodegradation:**

130 Removal of contaminants in a biological filter is a multi-step process involving adsorption,
131 biofilm formation, biodegradation, desorption and diffusion of contaminants and nutrients
132 across the biochar-biofilm-water interface ^{19,20}. **Fig. 1** illustrates the various steps,
133 mechanisms and processes that might occur during the dynamic biochar-biofilm interactions
134 for contaminant removal in BEB filters. Microbes generally attach themselves to the biochar
135 surface by secreting a gluey, Extracellular Polymeric Substances (EPS), made of
136 biopolymers^{18,25,27}. The transport of contaminants and other water-borne substances from
137 bulk fluid to this EPS is often dominated by molecular diffusion. Microbes then metabolise
138 these organic and inorganic contaminants via several biochemical and/or bio-electrochemical
139 reactions involved in respiration and cell growth ^{15,25,28}.



140

141 **Figure 1: Adsorption, biosorption and biodegradation in Biologically-Enhanced Biochar**

142 **filters** A) Diffusion and adsorption of target contaminants from bulk to biochar surface, B)

143 Microbes forming biofilm with Extracellular Polymeric Substances C) Diffusion of

144 contaminants through Extracellular Polymeric Substances D) Microbes metabolise target

145 contaminants, releasing simpler product molecules; biosorption based on biofilm's adsorption

146 affinity for contaminants/metabolic by-products, E) Spontaneous bio-filter bio-regeneration

147 via desorption of partially degraded contaminants and metabolic products owing to reverse

148 concentration gradients

149 Researchers have widely studied the adsorptive properties of biochar in water treatment,

150 however studies that investigate the combined adsorption and biological degradation of

151 contaminants using biochar is an emerging area of research^{10,29-32}. This use of 'Biologically-

152 Enhanced Biochar', despite a limited number of studies, has promising results for the

153 removal of organic/inorganic/biological substances from wastewater. The biofilm and its

154 biodegradation by-products are also capable of adsorption of several organic-inorganic

155 contaminants via biosorption (adsorption by biomass) ^{33,34}. Microbial biofilms and their
156 biosorption capabilities for bioremediation form a huge area of research ^{33,35}. For example,
157 biochar units adsorbed arsenic from wastewater followed by separate periphytic biofilm
158 reactors (containing both heterotrophic and phototrophic microbes), which adsorb the
159 remaining arsenic in the water via biosorption by biofilms ³⁶.

160 The contaminant removal efficiencies and mechanisms of different BEBs can vary
161 significantly depending on process conditions, type of contaminants and biochar properties.
162 For example, Aspen wood biochar showed a decrease in the degradation of naphthenic acids,
163 while softwood bark biochar showed a significant increase in naphthenic acid degradation;
164 both studied in the presence of metal contaminants³⁴. Both biochars supported biofilms of
165 similar thickness. Here, each biochar by virtue of their unique physical and chemical
166 properties, led to a different selection of microbial community. This different biofilm
167 proliferation will lead to different metabolic potential and explains the observed difference in
168 the degradation capacities. This rationale is reinforced by the finding that naphthenic acid
169 degradation capacities were comparable for both biochar types in sterile conditions.
170 Contaminant removal due to adsorption alone was close to 30% when tested on sterile
171 biochar samples compared to up to 87% with biofilm growth. A four-fold increase in metal
172 (Fe, Al, As) uptake from the water phase further confirms the influence of biosorption. This
173 points to other interplaying parameters, such as the physico-chemical properties of the
174 biochar support, microbial community structure and its metabolic potential, type of
175 contaminants, kinetics and mechanisms of biodegradation, which would all dictate the overall
176 contaminant removal capacity of a BEB filter.

177 Dalahmeh et. al. investigated and compared the degradation of four pharmaceutical
178 compounds (carbamazepine, metoprolol, ranitidine and caffeine) by hardwood biochar and

179 sand filters ³⁷. The study investigated biochar and sand filters with active, inactive and no
180 biofilms to quantify the effects of adsorption, biodegradation, and a combination of these
181 processes on contaminant removal. The results suggest that the extent of contaminant
182 removal varies greatly with the type of pharmaceutical contaminants, the mechanism of
183 contaminant removal and also the filter material used (biochar or sand). Biochar filters
184 performed better than sand filters for carbamazepine and metoprolol removal with adsorption
185 as the main mechanism of contaminant removal. Both biochar and sand filters had
186 comparable contaminant removal efficiencies for ranitidine and caffeine via a combination of
187 adsorption and biodegradation. Biochar filters with active biofilm performed better than sand
188 filters for the removal of organic matter and nitrogen. The study was extended for the
189 removal of per-and polyfluoroalkyl substances (PFA) using the same model to find that
190 biochar efficiently removed long-chain PFAs with chain length greater than C6³⁸. For similar
191 process conditions, the different solubility of contaminants in water, the adsorption affinities
192 of filter for various contaminants and organic matter, and the dissimilar biodegradability of
193 contaminants are the factors which have been identified to result in differences in the
194 efficiency of contaminant removal.

195 Apart from removing organic/inorganic contaminants, biochar-based water filters are also
196 efficient in removing microbial pathogens such as *Escherichia.coli*, *Staphylococcus aureus*
197 from contaminated water ³⁹⁻⁴¹. Biochar amendment of conventional sand/compost bio-filters
198 used for storm water treatment is another promising and emerging area of research ⁴². These
199 biochar-based interventions can be particularly useful in the light of extreme climate events
200 such as floods, especially for people in LMICs, who are usually the worst-affected.

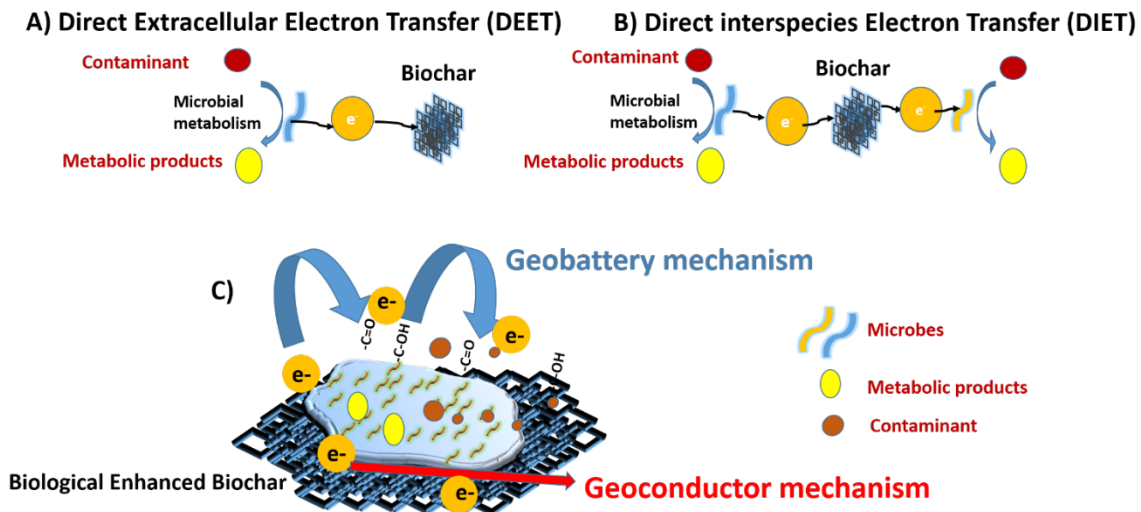
201 Studies on removal of *E. coli* in biochar-amended filters show how properties of biochar such
202 as surface area, polarity, particle size and the biofilm properties such as microbial geometries

203 have a very significant effect on capacity of *E. coli* removal^{40,43}. Biochar of small particle
204 size with low volatile matter content and low polarity were found to be more beneficial for
205 *E.coli* removal, while the infiltration rates and initial bacterial composition did not have a
206 large influence⁴⁰. The main reasons for the improved *E. coli* removal capacity upon addition
207 of biochar appear to be the improved water-holding capacity of biochar-amended bio-filters
208 and the higher attractive/binding forces of biochar surfaces. A recent meta-analysis relates
209 this improvement in soil structure with biochar amendment to an increased soil porosity,
210 number of pores and pore connectivity⁴⁴. This leads to better *E. coli* attachment on biochar
211 surfaces, increasing their overall removal from infiltrating storm water. The authors also
212 discuss the detrimental effects of increasing dissolved organic content in the system on *E.*
213 *coli* removal, although a definite explanation for this is yet to be found.

214 Afroz. et. al. evaluated the effects of biochar amendment in laboratory-scale bio-filters on
215 two bacterial pathogen removal efficiencies (*Salmonella enterica serovar Typhimurium* and
216 *S. aureus*), as well as bacterial and viral indicators *E. coli* and *MS2 coliphage*⁴⁵. Biochar
217 amendment resulted in a considerably higher increase in bacterial removal efficiency (9-fold
218 increase) compared to bacteriophage removal efficiency (3-fold increase). The study also
219 finds the experimentally observed microbe removal rates to be 2.8-7 times higher than the
220 theoretically calculated removal rates (using Colloidal Filtration Theory (CFT)). The
221 observed effects are attributed to removal mechanisms such as straining (a sieving effect,
222 where contaminants of large size cannot pass through the small pore size of biochar filter
223 medium) and hydrophobic interactions that are likely to be affected by biochar. These
224 mechanisms are specific to each type of microbe and are not included in CFT. This further
225 highlights the complexities of biochar/microbe interactions and demonstrates how the current
226 models are insufficient in studying these interactions.

227 **3.2 Bio-electrochemical interactions in BEB:**

228 Microbes often use redox active species such as organic matter, ammonium, O₂, NO₃⁻ etc. as
229 energy sources, energy sinks, energy storage materials, and in cell-to-cell electron
230 transfers^{46,47}. Biochar, being electrically conductive and with abundant redox active sites
231 (such as quinone, phenolic groups) can mediate and support microbial metabolism
232 effectively^{48,49}. In some cases, the porous carbon support may just act as ideal supports,
233 allowing a good biofilm growth, but not actively taking part in the degradation⁵⁰. While a lot
234 of metabolic biochemical reactions in microbes are intracellular, certain microbes are capable
235 of directly transferring electrons between intracellular redox moieties and external redox sites
236 such as minerals and electrically conducting carbon⁵¹. Such microbes capable of extracellular
237 electron transfer have been identified as 'electroactive microorganisms'^{28,47,52}. The mere
238 presence of electroactive substrates has been shown to improve microbial biodegradation⁵³. It
239 is thus highly likely that an immobilized biofilm will proliferate in a way that makes use of
240 the electrical conductivity and redox sites in biochar for bioremediation^{11,49}. Biochar can also
241 be carriers of humic substances and redox mediators such as metal composites, which provide
242 additional electron exchange capability for mediation in microbial metabolism^{48,54}. This use
243 of electroactive biochar for energy and environmental applications using Microbial
244 Electrochemical Technologies (METs), is another emerging area of research^{11,49,55}. Biochar-
245 based METs can make use of microbial catalysis in electrochemical processes and can
246 valorise several forms of waste (solid, liquid, and gas) for sustainable generation of a wide
247 range of products such as electricity, biofuel and biogas^{55,56}.



248

249 **Figure 2 Bio-electrochemical interactions in Biologically-Enhanced Biochar filters A)**

250 Direct Extracellular Electron Transfer (DEET) between electroactive microbes and biochar

251 via membrane-bound cytochromes or conductive nanowires manifested in the cells, B) Direct

252 interspecies Electron Transfer (DIET) between two different electroactive microbe species,

253 biochar being both electron acceptor/donor, C) Both DIET and DEET can happen via:

254 Geobattery mechanism- Electron shuttled through the various electron donating/accepting

255 groups $-C-OH$, $C=O$ or Geo-conductor mechanism- electrons shuttled through the conductive

256 graphitic frameworks of biochar

257 Electroactive microbes can metabolise contaminants via Direct Extracellular Electron

258 Transfer (DEET) and Direct Interspecies Electron Transfer (DIET) ^{55,57}. In DEET,

259 microorganisms physically connect to biochar for electron transfer (**Fig. 2**) ^{13,48,58}. DEET also

260 occurs when microbes utilise nanowires or pili (vesicular extensions of periplasm or outer

261 cell membranes, 2-3 μm long) to connect to the electrodes, especially for interconnections

262 with electrode surfaces from within the deeper layers of the biofilm⁵⁹. In DIET,

263 microorganisms utilise other cells as electron mediators via a syntrophic metabolism²⁸.

264 Electrically conducting biochar can promote DIET significantly in processes such as

265 wastewater treatment and anaerobic digestion ^{14,60,61}. For both DEET and DIET, one way of

266 electron transfer is through the highly conducting graphitic frameworks of biochar, often
267 referred to as the geoconductor mechanism^{11,62} (**Fig. 2**). It is also possible to shuttle electrons
268 by the interaction of surface functional groups such as quinones (electron accepting) and/or
269 phenolic/hydroquinone (electron donating) groups by reversibly accepting and donating
270 electrons, referred to as the geobattery or geocapacitor mechanism^{11,62}.

271 Prado et. al. found maximum biodegradation efficiencies by electro-conductive biochar in
272 both batch and continuous operations, compared to coke and graphite, despite graphite
273 showing 40,000 times higher electrical conductivity than biochar⁵³. Biochar displayed a
274 larger number of electroactive functional groups compared to coke and graphite. If the
275 geoconductor mechanism was dominant, graphite would be outperforming biochar in
276 biodegradation via geoconductor mechanism, which is not the case. The study thus pinpoints
277 to a possible geobattery mechanism of electron transfer using the abundant functional groups
278 in biochar, which would have enabled it to outperform graphite and coke. However, we point
279 out that the biochar used here had a higher surface area (almost 200 times more) and total
280 pore volume (100 times higher), than the graphite and coke used. It would be interesting to
281 see if biochar with a similar surface area and pore size distribution to coke and graphite
282 would perform well under the same conditions.

283 The ability of biochar to promote DIET in anaerobic digestion is a very recent discovery and
284 not fully understood yet^{12,63}. Even less is known about the role of biochar DIET mediation in
285 biological water treatment. However, due to the prevalence of microbial metabolic activities
286 and biochar-biofilm interactions, it is highly likely that DIET occurs in biological water
287 treatment as well. Previous studies confirmed DIET as the mechanism of syntrophic
288 metabolism, via pili (conductive nanowires) formation in *Geobacter. metallireducens*⁶⁰.
289 However, the process of pili production is slow, and thus, presence of a conductive material,

290 such as biochar can facilitate the electron exchange process by acting as an electrical conduit,
291 by-passing the mechanism relying on pili, thus making the entire metabolic process much
292 faster. A biochar-amended co-culture in a recent study showed a 15x higher ethanol
293 metabolization rate in presence of biochar compared to a co-culture with no biochar⁶⁰. The
294 metabolization rates were comparable to rates reported by Granular Activated Carbon, which
295 had 1000x more electrical conductivity than biochar⁶⁰. However, no direct link between the
296 rates of ethanol metabolism and electrical conductivity of biochar could be established. This
297 indicates that a number of other additional factors such as porosity, aromaticity, and electron
298 exchange capacities may play an important role in DIET. **Table 1** provides more details on
299 the process and experimental conditions for the examples discussed under Section 3.

300 While it is clear that biochar can support and mediate microbial metabolism leading to
301 removal/degradation of water-borne contaminants, there is no clear consensus on which
302 process conditions and properties of BEB promote particular mechanisms. We also do not
303 know how, why and to what extent each of these mechanisms discussed above can contribute
304 to contaminant removal under a certain set of conditions. Future research should focus on
305 decoding these complex and dynamic interactions of BEBs to be able to systematically
306 engineer BEBs for biological water treatment.

307

Table 1- Experimental details of Biologically-Enhanced Biochar studies discussed in Section 3

Biochar type and pyrolysis conditions	Biochar properties	Contaminant/Influent	Contaminant removal mechanisms and observations	Process conditions	Ref.
<p>Bismuth-impregnated Wheat straw</p> <p>Production conditions: HTT=500 °C, HR = 10°C min⁻¹, Residence time= 60 min</p>	<p>SSA: 190.4 m² g⁻¹,</p> <p>Avg. pore dia.= 2 nm</p>	<p>Target contaminant: As (III)</p>	<ul style="list-style-type: none"> • Adsorption by biochar and biosorption by periphytic biofilm, Total As (III) removal rate (90.2–95.4%). • Adsorption removes 60 % As (III) and remaining As by periphytic reactors via calcite, OH and CO groups in biofilm 	<ul style="list-style-type: none"> • Biochar column reactor (565 mL, dia: 6 cm, height: 20 cm), spiral periphyton bioreactor (21.9 m: length, dia: 2.0 mm diameter) • Influent flow rate of = 1.0 mL min⁻¹ • Initial As (III) concentrations of 0, 2.0, 5.0, 10.0 and 15.0 mg L⁻¹. 	36
<p>Aspen wood (N3) and Softwood (SB)</p> <p>Pyrolysis conditions: n.a</p>	<p>SSA: 4 m²g⁻¹ (N3), 189m² g⁻¹ (SB)</p> <p>Total pore volume: <0.01 mL g⁻¹ (N3), 0.12 mL g⁻¹ (SB)</p>	<p>Influent: Oil sand process water (OSPW)</p> <p>Target contaminants: Naphthenic acids (NA), Fe, Al, As</p>	<p>Adsorption, Biosorption, Biodegradation</p> <ul style="list-style-type: none"> • N3 and SB best-performing among 8 carbon supports; • Biodegradation higher for SB-associated biofilms with NA removal at 87% in presence and 72 % in absence of Fe, Al and As metals • Biosorption enabled up to four times more removal of Fe, Al, and As 	<ul style="list-style-type: none"> • Microbial-biochar attachment assays using 24-well cell culture plates • Initial NA concentration of 200 mg L⁻¹ 	34
<p>Hardwood (mixture of pine-spruce wood)</p> <p>Pyrolysis conditions: HTT= 800 °C</p>	<p>SSA: 184 m² g⁻¹</p>	<p>Influent media: Municipal wastewater</p> <p>Target contaminants: Carbamazepine, Metoprolol, Ranitidine, and Caffeine, Per-and polyfluoroalkyl substances (PFASs)</p>	<p>Adsorption, Biosorption, Biodegradation-</p> <ul style="list-style-type: none"> • Carbamazepine and metoprolol removal- Biochar> sand-adsorption is dominant. • Ranitidine and caffeine removal- Biochar>Sand-combination of adsorption and biodegradation • Organic matter and nitrogen removal- Biochar with active biofilm> sand filters 	<ul style="list-style-type: none"> • Column filters (diameter 5 cm; height 55 cm) used over a period of 22 weeks <p>Contaminant concentration ranges:</p> <ul style="list-style-type: none"> • 5400–25000 ng L⁻¹ for carbamazepine, • 260-39000 ng L⁻¹ for metoprolol, • 2000–10300 ng L⁻¹ for ranitidine, • 1800-11000 ng L⁻¹ for caffeine • 1500-4900 ng L⁻¹ for PFASs 	37, 38
<p>Softwood</p> <p>Pyrolysis conditions: HTT= 815-1315 °C. Residence time=1 to 3 s, Used as Biochar-augmented sand filters</p>	<p>Composition: 79% carbon, 12% ash 16% volatile matter</p>	<p>Influent: Synthetic storm water</p> <p>Target contaminant: E.coli</p>	<ul style="list-style-type: none"> • Small biochar particle size with low volatile matter content and low polarity, low Dissolved Organic Content in media better for E.coli removal • Infiltration rates and initial bacterial composition do not affect E.coli removal • Up to 96 % E.coli removal rate in biochar-augmented sand filters 	<ul style="list-style-type: none"> • Glass chromatography column filters (Kontes, 15 cm length, 2.5 cm diameter) with Teflon fittings at both ends with a built-in mesh (20-µm pore opening) • Initial E.coli concentration- 10³ to 10⁷ colony forming units (CFU) mL⁻¹, infiltration rate constant at 12 cm h⁻¹ 	40

<p>Feedstock: n.a (commercial biochar)</p> <p>Pyrolysis conditions: n.a</p>	<p>SSA: 104.64±7.80 m² g⁻¹</p>	<p>Target contaminant: Bacterial pathogens: Salmonella enterica serovar Typhimurium and Staphylococcus aureus, Bacterial and viral indicators: Escherichia coli and MS2 coliphage</p>	<ul style="list-style-type: none"> • Electrostatic interactions in bacteriophage removal, Straining and hydrophobic interaction in bacterial removal. • Up to 3.9, 1.9, and 1.8 log₁₀ removal for pathogenic bacteria, E. coli, and MS2, respectively. 	<ul style="list-style-type: none"> • Polyvinyl chloride (PVC) pipes (2.5 × 15 cm) reactors with end fittings and glass wool at both ends • Volumetric flow rate of 1 mL min⁻¹ • E.coli and staphylococcus concentration- ~10⁵ (CFU) per mL, Salmonella concentration ~10³ CFU per mL, MS2 ~10¹¹ Plaque Forming Units (PFU) mL⁻¹ concentrations 	<p>45</p>
<p>Quercus wood (commercial biochar)</p> <p>Pyrolysis conditions: n.a</p>	<p>SSA: 210-250 m²g⁻¹</p> <p>Total pore volume 0.12-0.13 cm³g⁻¹</p>	<p>Influent: Synthetic and Urban wastewater</p> <p>Target contaminant: Organic content</p>	<ul style="list-style-type: none"> • Bio-electrochemical interactions- Biochar outperformed graphite and coke via geobattery mechanisms, • Maximum removal efficiency (92%) and degradation rate (185 g-COD m³d⁻¹) at anodic potential as high as 0.6 V 	<ul style="list-style-type: none"> • Snorkel bio-filters 24 cm high and 3 cm internal dia., total bed volume of 170 cm³ and a hydraulic volume of 100 mL • Separate three-electrode configuration for electrochemical measurements • Real urban wastewater treated at two different organic loading rates (OLR- 170 mg L⁻¹ and 890 mg L⁻¹), two different hydraulic retention times (HRT- 4 and 2 days) and three different anode potentials (short circuit (non-polarized), 0.4 V and 0.6 V vs. Ag/ AgCl) 	<p>53</p>
<p>Pine wood biochar ESI, BEC and Kiln biochar (KBC)</p> <p>Pyrolysis conditions: ESI- HTT: 500°C for 2 hours BEC- HTT: 700 °C for 30s and 500°C for 15 min KBC - HTT 600°C for 2 hours</p>	<p>SSA: ESI - 167 m² g⁻¹ BEC - 15 m² g⁻¹ KBC - 209 m² g⁻¹</p> <p>Electrical conductivity: ESI - 2.1 μS cm⁻¹ BEC - 4.4 μS cm⁻¹ KBC - 4.3 μS cm⁻¹</p>	<p>Biochar mediation and impact on syntrophic associations in ethanol and fumarate media</p>	<p>Direct Interspecies Electron Transfer</p> <ul style="list-style-type: none"> • Co-cultures of Geobacter metallireducens with Geobacter sulfurreducens (succinate production) or Methanosarcina barkeri (methane production) amended with biochar stimulated syntrophic association leading to higher metabolism of ethanol compared to samples not amended with biochar • Scanning Electron Microscopy suggests electrical connections between the species were through biochar rather than via cell-to-cell electron transfer 	<ul style="list-style-type: none"> • Pure cultures and co-cultures were incubated anaerobically, in 27 mL pressure tubes with 10 mL medium under an anoxic atmosphere of 80:20 of N₂:CO₂ 	<p>60</p>

*HTT- Highest treatment Temperature, HR- Heating Rate, SSA- Specific Surface Area, Avg. pore dia.- Average Pore Diameter

308 **4. Design of BEBs**

309 Understanding contaminant removal mechanisms and biochar-biofilm interactions is
310 paramount to design of effective and efficient BEB filters. Producing the right biochar tuned
311 for a specific application, i.e., supporting selected contaminant removal mechanisms (such as
312 adsorption, biodegradation, biosorption, bio-electrochemical interactions) and targeting
313 specific contaminants is of utmost importance for it to be a viable option in water treatment.

314 Surface area is usually considered to be an important prerequisite for supporting biofilms and
315 contaminant removal. However, there is increasing evidence suggesting that a high surface
316 area alone cannot guarantee high performance of porous-carbon filters, especially where
317 several contaminants are present and multiple dynamic mechanisms in action^{34,64,65}. Most
318 microbes measure around 1-2 μm in size and hence cannot directly access micropores (< 2
319 nm), mesopores (2-50 nm), or macropores (50-200 nm) present in biochar and activated
320 carbon^{50,66}. However, while not best suited for direct biofilm formation, the adsorption of
321 nutrients or contaminants in these micro- and mesopores can provide improved conditions for
322 microbial colonisation in adjacent larger pores and interparticle surfaces. At these external
323 surfaces, fluid flow and mass transfer effects play a deciding role in biofilm formation and
324 geometry⁶⁷. Activated carbons displaying high surface areas consist mostly of micropores,
325 making more meso- or macro-porous material such as biochar potentially more suitable for
326 biofilm formation^{50,66}. However, the dynamic attachment and growth of microorganisms
327 forming biofilms in three-dimensional porous media such as biochar is not yet fully
328 understood. Thus, an important aspect of BEB design will be to study the effects of biochar
329 pore structure on biofilm establishment and biodegradation. There is growing research in this
330 field to understand and discern this inter-dependence of porous media characteristics (pore
331 size, pore distribution, pore connectivity, roughness, geometry, hydrophobicity-

332 hydrophilicity), mass-transfer processes, biofilm proliferation (geometry,
333 homogeneous/heterogeneous distribution of biofilm in porous media), wastewater properties
334 (physico-chemical and biological properties) and cell surface properties ^{67,68}. This will also
335 help to understand how the competing forces of pore clogging (from the growing biofilm,
336 biodegradation products and contaminants) and biofilm growth and geometry (which needs
337 space for growth and free diffusion of nutrients, oxygen) influence biodegradation, eventually
338 enabling the design of more efficient BEB filters.

339 Using a biofilm-biochar combination without first considering its specific function may not
340 help in removal/degradation of contaminants in wastewater. The choice of each component
341 and their properties as well as the composition of influent water and operating conditions
342 have to be taken into account. The interplay of these numerous parameters determines
343 whether the presence of a biofilm on biochar enhances or inhibits the rate and extent of
344 contaminant removal. Characterisation techniques and analytical tools suitable to study the
345 complexities of BEB systems are provided in **Table 2**. Control parameters and process
346 conditions which can be used in fine tuning these properties, are shown in **Table 3**. Even
347 though these are properties that have shown to have profound effects on water-borne
348 contaminant removal, additional studies focussing on correlating these properties to their
349 process conditions and BEB properties are necessary to optimise BEBs effectively.

350 Studying BEB filters with model-based computational tools would be one way to better
351 understand how these various biochar-biofilm properties, process conditions and contaminant
352 efficiency are interdependent. Although a lot of mathematical models and statistical analyses
353 have been used to study biochar properties, microbial biofilms and biofilm reactors
354 individually, studies that integrate all the complexities of biological filters (biochar and
355 biofilm) on a fundamental level are still largely missing in literature^{2,53-56}.

356 **Table 2** Techniques and analytical tools for BEB characterisation

Techniques and tools	Information
Compositional analysis	
TGA	% (Fixed carbon , volatile matter, ash, moisture)
Elemental analysis	CHNSO composition, biochar stability
ICP-OES/MS, XRF	Mineralogical and elemental composition
XAS (XANES, EXAFS)	Local geometric and/or electronic structure
Surface properties	
Surface profilometer	Topographical analysis
Surface charge analyser	Zeta potential
Contact angle goniometer	Surface wettability
FTIR, Boehm titration	Surface functional groups
XPS	Surface chemistry, bonding information
Morphology and structure	
SEM/EDX	Surface morphology, elemental mapping
XRD, Raman spectroscopy	Crystallinity/amorphous nature
Gas physisorption using N ₂ , Ar, H ₂	Pore-size distribution, surface area
X-ray μ -tomography	Internal 3D structure, pore-connectivity
Electrical and Electrochemical properties	
CV, EIS, other amperometry/potentiometry	Bio-electrochemical interactions, mechanisms
4-point probe resistivity, EIS	Electrical conductivity
Influent/effluent composition, BEB toxicity	
HPLC, LC-MS, GC-MS	Contaminant composition, concentration
Biofilm properties	
16rDNA illumina sequencing, RISE	Biofilm composition-function
SEM, Confocal/Fluorescence microscopy, FISH	Biofilm growth, structure, diversity

357 TGA- Thermogravimetric analysis, ICP-OES/MS- Inductively Coupled Plasma- Optical Emission
 358 Spectroscopy/Mass Spectrometry, XRF- X-ray Fluorescence, XAS-X-ray Absorption Spectroscopy,
 359 EXAFS- Extended X-ray Absorption Fine Structure, XANES- X-ray Absorption Near Edge Structure

360 (XANES), FTIR- Fourier Transform Infrared Spectroscopy, XPS- X-ray Photoelectron Spectroscopy,
 361 SEM/EDX- Scanning Electron Microscopy/Energy Dispersive X-ray spectroscopy, XRD- X-ray
 362 Diffraction, CV- Cyclic Voltammetry, EIS- Electrochemical Impedance Spectroscopy, HPLC- High
 363 Performance Liquid Chromatography, LC-MS- Liquid Chromatography- Mass Spectrometry, GC-MS-
 364 Gas Chromatography- Mass Spectrometry, RISE- rRNA intergenic spacer analysis, FISH- Fluorescence
 365 In situ Hybridisation

366 **Table 3** Process conditions and biochar production (pyrolysis) conditions crucial for
 367 optimising BEB properties, based on literature data

Process conditions	Pyrolysis conditions
Influent concentration, Dissolved Oxygen, Highest Treatment Temperature (HTT), Particle size, pH, Upstream/downstream treatments, Empty bed contact time, Post-treatment	Biomass feedstock, Pre-treatment steps, Temperature, Pressure, Carrier gas, Gas flow rate, Heating/cooling rate

368

369 Experimental findings can be used to complement and improve these models and promote a
 370 fundamental understanding of how biological filters work under different conditions. Smolin
 371 et.al. developed a model to quantify adsorption, biodegradation, and self-bio-regeneration in
 372 BAC for organic contaminants²⁷. The model allowed a quantitative assessment of cooperative
 373 removal (adsorption and biodegradation) of organic matter by taking into account the non-
 374 target filling of BAC pores with products of metabolic activity, showcasing potential insights
 375 from advanced modelling. This model was applied to evaluate and study the biological
 376 filtration of 2-nitrophenol through a BAC filtration unit for 38 months and found that the
 377 activated carbon adsorption efficiency was preserved for almost three years as a consequence
 378 of bio-regeneration. It was also found that there was an increased non-target porosity loss
 379 (metabolic by-products filling pores) with increase in operation time with an estimated
 380 porosity loss of 61% of pore volume of fresh sorbent at the end of the experiment.

381 Correlating changes in microbial community structures and function with changes in biochar
 382 properties is an important area requiring more attention as it can help in identifying BEB

383 properties which impact biodegradation capacities. Due to recent technological advancements
384 in molecular biology and data science, there is a fast-growing body of literature on microbial
385 community structures and function in water treatment plants⁶⁹⁻⁷¹.

386 Dalahmeh et. al. found biochar filters to have largely similar microbial community structures
387 and diversity compared to sand filters, but an enhanced diversity in α -proteobacteria and γ -
388 proteobacteria in biochar filters resulted in a larger biodegradation potential for organics³⁷.

389 The experiments were performed in column filters of height 55 cm and diameter 5 cm.

390 Additionally, a change in filter media (bark, charcoal and sand filters) for the same process
391 conditions resulted in colonisation by different genera of biofilm-forming bacteria with
392 community structure and potential respiration rate, also changing over time and filter-bed
393 height⁷². It was found that most of the organic biodegradation happened in the top 20 cm for

394 bark and charcoal filters, allowing filter-beds to be shallower compared to sand filters.

395 Similar studies investigated changes in biofilm composition for biological activated carbon
396 supports with different surface chemistry for 4-nitrophenol biotransformation in anaerobic

397 conditons⁵⁰. The results suggest that even though activated carbon cloths with different
398 surface properties induced changes in microbial community structures in the resulting

399 biofilm, the biotransformation efficiencies of 4-nitrophenol for these different types of carbon

400 supports were very similar, i.e. close to 59 %. The microbial composition indicated an

401 enrichment of exoelectrogenic microorganisms (*Geobacter* spp.) over activated carbon cloth

402 surface compared to controls with no support material, as revealed by 16S rRNA amplicon

403 sequencing. These examples reiterate the complexities of biodegradation and the need for

404 more fundamental research decoding these dynamic biochar-microbial interactions.

405 **5. Opportunities for BEBs in water treatment systems**

406 Activated carbon, used for Biological Activated Carbon process, is a benchmark material
407 largely used in tertiary water treatment applications. While biochar is mainly produced from
408 renewable biomass and organic residues, a large fraction of AC production uses non-
409 renewable sources such as coal, a large and cheap carbon source^{73,74}. Water treatment and air
410 purification applications alone account for over 77% of the total global AC production⁷⁴.
411 Compared to biochar production, conventional AC production has additional energy
412 requirement which stems from the high-temperature/chemical treatment steps to activate the
413 surface and increase its surface area^{75,76}. After use, the exhausted-activated carbon is replaced
414 or regenerated using thermal treatment or steam activation before further reuse⁷⁷.
415 Regeneration is expensive and energy-intensive, accounting for approx. 30% CO₂ equiv.
416 compared to the virgin AC⁷⁷⁻⁷⁹. These limitations of AC restrict its widespread use in
417 LMICs, despite its proven efficacy in water treatment applications. This urges us to explore
418 more sustainable options such as BEB for effective water treatment and associated end-of-life
419 treatments.

420 Countries like India, China, and Indonesia burn agricultural crop residue in the open causing
421 severe air pollution⁸⁰⁻⁸². India emitted 211 Tg of CO₂-equivalent GHG emissions in 2017 by
422 burning crop residue alone⁸³. Developing biochar-based materials for sustainable water
423 treatment can thus greatly benefit countries with large agrarian sectors. Community-based
424 biochar-based water treatment solutions, ranging from home-scale to large-scale will improve
425 access to potable water while mitigating the impact of burning biomass in the open. There are
426 recent and encouraging studies which demonstrate biochar production using traditional kilns
427 such as 'Kon-Tiki' kilns, especially in rural settings^{84,85}. This will mean that there are more
428 sources of potable water, while also ensuring lesser greenhouse gas emissions compared to
429 burning biomass in the open^{84,85}. Thus, BEB not only becomes an important material that
430 could become a cheaper and sustainable substitute for BAC in water treatment, but also offers

431 an additional advantage of being a stable carbon sink having the potential to offset the CO₂
432 emissions from fossil fuels^{4,86,87}.

433 **5.1 Economic and environmental considerations**

434 To further explore the potential of biochar as a sustainable candidate in biological water
435 treatment, we use published Life Cycle Assessments (LCA) and meta-analysis from literature
436 to compare biochar with activated carbon, a benchmark in tertiary water treatment processes.
437 Since LCAs specific to biological water treatment could not be found in literature, we draw
438 these comparisons using studies which have used a wide range of biochar and AC products,
439 currently used in wastewater treatment. We also point out that possible differences may arise
440 from the interplay of biological mechanisms, which are not accounted for in the selected
441 studies. Special caution is necessary in drawing generalised conclusions from LCAs on
442 specific biochars due to differences in production conditions as well as feedstock sources and
443 therefore biochar properties.

444 Thompson et al. quantified environmental impacts of biochar produced from wood and
445 biosolids and compared it to various coal-derived AC for use as adsorbents for removal of
446 sulfamethoxazole (SMX), a common antibiotic found in wastewater⁷⁶. The assessment of
447 biochar and AC capacities (low and moderate) was based on the ability of adsorbents to
448 remove SMX, determined as the adsorbent dose required to achieve a 75% SMX adsorption
449 from wastewater effluent following 60 min contact. The results showed that moderate
450 capacity wood biochar had the lowest overall environmental impact, outperforming the other
451 materials on 8 environmental criteria out of the 10 considered. This was largely due to the
452 carbon sequestration potential of biochar and the energy production during pyrolysis of wood
453 biomass. This shows that under specific circumstances, biochar can outperform AC in terms
454 of environmental benefits despite having a lower specific capacity for contaminant removal.

455 However, biosolid-based biochar had a higher environmental impact compared to both AC
456 and wood-derived biochar due to the high energy requirements in drying biosolids prior to
457 pyrolysis. Here, the specific capacity of biochar to remove SMX was approximately 8.5 times
458 lower than that of activated carbon. Contrarily, when specific capacity of biochar 40 times
459 lower than AC was used in the analysis, activated carbon became an environmentally
460 preferred option⁸⁸. This indicates that not all types of biochar are suitable replacements for
461 activated carbon and conclusions have to be drawn on a case by case basis. It would also be
462 useful to compare sustainability parameters of activated to non-activated biochar, systematic
463 studies of which are currently missing in literature. However, it is important to point out that
464 the contaminant removal in these LCA studies were purely based on adsorption, where
465 biochar has the disadvantage of a lower surface area. The relative performance of biochar and
466 activated carbon is different for biological water treatment, where biofilms regenerate
467 adsorption capacity of the surface and total surface area is less important, and other properties
468 of biochar such as a pore-size distribution may play a more prominent role, making BEBs
469 competitive on performance along with environmental impact.

470 Results of a meta-analysis study comparing 80 different types of biochar and AC for heavy
471 metal removal showed that biochar had lower energy demands and lower Global Warming
472 Potential (GWP) compared to activated carbon⁷⁵. The average energy demand for production
473 of biochar was considerably lower, at 6.1 MJ/kg, than that for AC, at 97 MJ/kg, mainly due to
474 the energy intensive process of activation and relatively low AC yields, especially when
475 produced from renewable feedstock. Consequently, the average GWP of biochar production
476 was negative at -0.9 kg CO₂-eq/kg due to the carbon-sequestration potential of biochar, while
477 AC had an average GWP of 6.6 kg CO₂-eq/kg⁸⁹⁻⁹¹.

478 In addition to the assessment of environmental performance of biochar vs AC as a filter
479 material, the economic perspective is also important to consider. One way to evaluate the
480 economic performance is to compare the cost of removing a certain mass of contaminants.
481 For example, the cost of using biochar as an adsorbent is lower/equal in the case of Ca, Cr,
482 Cd, Zn as compared to AC, with the exception of lead⁷⁵. Average prices cannot reflect the
483 different capacities/efficiencies of biochar materials. This is indicated by the wide price range
484 for biochar from 200-1800 \$t⁻¹ as well as for activated carbon from 1100 - 5000 \$t⁻¹ ^{7,75,76,92}.
485 These findings highlight that both environmental and economic performance of biochar will
486 greatly depend on how well the properties of biochar are tuned to suit the final application.
487 Careful evaluation of the water treatment conditions, mechanisms of contaminant removal,
488 properties of contaminants and filter material are crucial in engineering such sustainable
489 water treatment units. Due to a lack of standard production conditions and understanding of
490 contaminant removal mechanisms, comparative economic assessments for biochar and
491 activated carbon are still greatly missing in literature.

492 **5.2 End-of-life for BEBs: Sequential biochar systems**

493 Besides efficiently designing BEBs, research needs to develop novel end-of-life use
494 techniques to reduce any environmental impact. Although the cost of biochar is considerably
495 lower in most cases than that of AC, it is still not negligible, especially for LMICs. Here, we
496 discuss a novel approach to lower the overall cost of biochar in wastewater treatment by
497 considering the whole value chain of the material over the full life cycle.

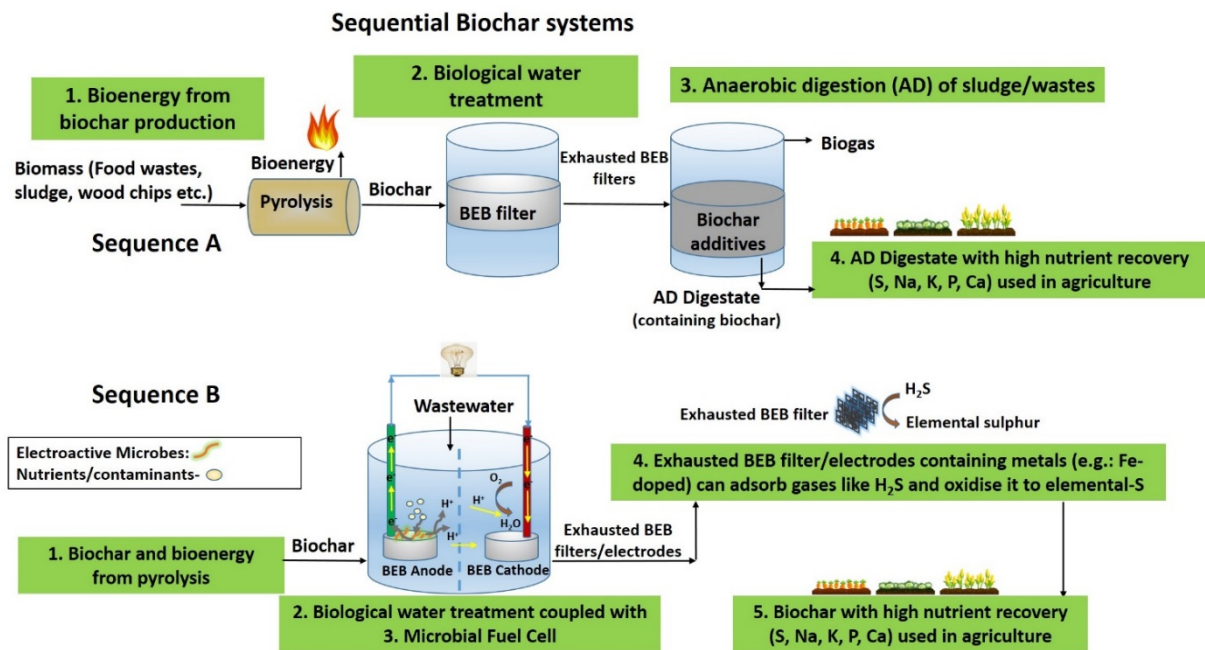
498 The concept of sequential biochar systems proposes utilising the multi-functional nature of
499 biochar to integrate different applications into the life cycle of biochar¹⁹. Due to the diverse
500 applications and variable properties of biochar, it can be economically and environmentally
501 advantageous to recycle biochar and use it for more than one application⁹³. As biomass

502 feedstock supply and biochar production are the two highest contributors to the economic
503 costs of biochar, multiple uses of biochar in sequence would offer the benefit of additional
504 revenue streams as well as splitting the production costs and environmental impacts over
505 several use phases⁵⁹. One of the simplest and best-tested approaches is the use of exhausted
506 biochar filter material as a fertiliser in agriculture. This enables the recycling of key nutrients,
507 such as phosphorous and nitrogen from wastewater back to food production⁹⁴⁻⁹⁶. However,
508 using exhausted filters for other applications such as agriculture should be done with caution
509 and only after a thorough safety assessment on a case by case basis. This is to avoid any risks
510 associated with potential leaching of toxic molecules (inorganic, organic and biological)
511 incorporated in biochar pores during its prior use in the water treatment process.

512 More complex sequences are currently under development, and we will discuss two examples
513 here. One feature that all the sequences have in common is the need for the biochar from one
514 application to meet the requirements of the next application. This can be achieved naturally
515 by appropriate sequence of use steps, or by adding an intermediate step to modify biochar
516 properties according to the requirements of the subsequent application.

517 In the first example, we propose a sequential use of biochar involving biological water
518 treatment and anaerobic digestion (Sequence A in **Fig. 3**). Here, BEB filters are used in
519 biological water treatment in step 2 after pyrolysis. The pyrolysis gas produced in Step 1 in
520 both sequences A and B (**Fig. 3**), could be used to partly provide for other energy
521 requirements of the sequence, such as biomass drying or as source of process heat. Biochar is
522 being increasingly used as an additive in anaerobic digestion to increase biogas production
523 ^{97,98}. Hence, we propose the use of exhausted BEB directly or with physical modifications
524 required to improve biogas production in anaerobic digesters, **Fig. 3**, step 3. The digestate,
525 containing biochar and other nutrients recovered from steps 2 and 3, can be further used in

526 agriculture (Step 4) ^{99,100}. This sequence might be especially relevant for agrarian
 527 communities due to the integration of several spatially related processes within industrial
 528 farm settings.



529

530 **Figure 3 Sequential biochar systems** Two examples of sequential biochar systems:
 531 Sequence A) Biologically-Enhanced Biochar (BEB) used in wastewater treatment, followed
 532 by the use of exhausted-BEB as additive in anaerobic digestion, and the use of digestate
 533 containing BEB in agriculture. Sequence B) BEB used in wastewater treatment, coupled with
 534 a Microbial Fuel Cell (MFC) for electricity generation using wastewater as the fuel,
 535 exhausted-biochar subsequently used in gas adsorption and soil application.

536 A more advanced example of a sequential biochar system is explained in Sequence B. The
 537 electro activity of biochar and electroactive microbes enables its use in Microbial
 538 Electrochemical Technologies (METs) such as Microbial Fuel cells for electricity generation
 539 and biological water treatment ^{49,56,101}. Biochar can act as a microbial inoculum carrier and
 540 can be used as electrodes (anode and cathode) of a microbial fuel cell, where respiring
 541 microbes convert chemical energy to electrical energy ¹⁰¹. Contaminated water can be used as

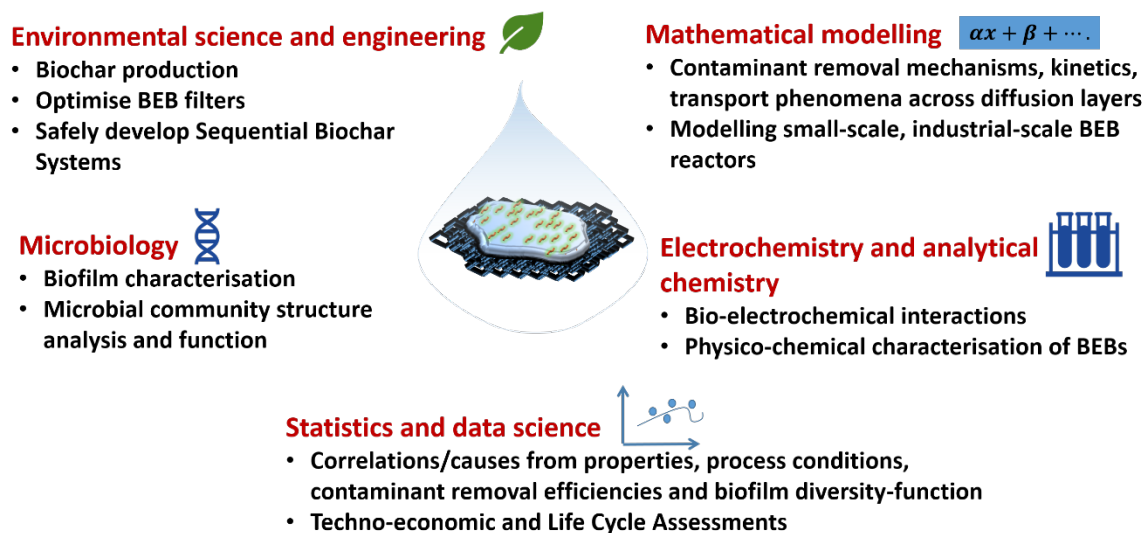
542 fuel, where microbes feed on the contaminants in wastewater. This makes them electricity
543 generators as well as biological water treatment units (**Fig. 3**, Sequence B, 2 and 3).
544 Electroactive microbes and the amount of energy they can produce still requires more in-
545 depth study, hence, use of METs are still challenging and in its nascent phase of large-scale
546 development²⁸. Biochar works as a strong adsorbent for gases and has the property to filter
547 and convert harmful gases such as H₂S and convert them in to less harmful materials like
548 elemental sulphur¹⁰²⁻¹⁰⁴ The exhausted biochar electrodes from MFC/water treatment units
549 can be used with/without additional treatments for use in end applications like gas
550 adsorption/removal **Fig. 3**, step 4)¹⁰²⁻¹⁰⁵. The biochar, with a high nutrient recovery from
551 steps 2, 3 and 4 can then be effectively used as soil additives in agriculture^{13,94} (**Fig. 3**, step
552 5).

553 **6. Challenges and Outlook**

554 Where water treatment with BEB is used as part of a sequence of different treatments,
555 successful establishment of biofilm and its growth during the start-up phase is important. The
556 effects of other upstream treatments such as coagulation, ozonation or chlorination on
557 bacterial growth and density have to be considered, as these treatments can impact biofilm
558 development. For example, residual chlorine in the influent water to a pilot-scale BAC unit
559 reduced the bacterial attachment on its surface, while a pre-ozonation process before BAC
560 treatment improved the biological activity of the biofilm for contaminant degradation in
561 many cases^{18,23,106-108}. These aspects should be considered during design of BEB filters for
562 industrial/ municipal applications.

563 It is important to design filters with pre-specified surface modifications to account for the
564 increasing number and concentration of contaminants in water, including Natural Organic
565 Matter (NOM), Contaminants of Emerging Concern (CECs) such as pharmaceuticals and by-

566 products of biodegradation. This is because high concentration of NOM and variations in
 567 nutrition can significantly alter the biofilm growth and efficiencies of biological filters^{22,109}.
 568 When exhausted filters are used, large amounts of organic matter and nutrients such as
 569 ammonia and phosphorus in the effluent water can lead to biofilm formation in the
 570 distribution networks^{18,26}. It is thus important to adjust the process parameters, frequency and
 571 intervals of backwashing and regeneration/change to fresh BEB filters¹⁰⁶. Competitive
 572 adsorption between target contaminants, non-target compounds such as organics and
 573 microbial metabolic by-products are a cause of concern for maintaining efficiency of
 574 biological filters. Research focusing on the fate and transport of transformation by-products
 575 formed during the biofilm metabolism of target contaminants is needed to investigate toxicity
 576 risks . The BEB designs should also account for biosorption of these microbial metabolic by-
 577 products. We identify several research areas requiring further attention to enable efficient
 578 design of BEBs, as illustrated by **Figure 4**.



579
 580 **Figure 4 Roadmap to Biologically-Enhanced Biochar (BEB) engineering** Schematic
 581 identifying the major areas of research vital for systematically engineering BEBs

582 The presence of toxic and carcinogenic compounds such as Polycyclic Aromatic
583 Hydrocarbons (PAH) and Volatile Organic Compounds (VOCs) that form during pyrolysis
584 and deposit on biochar surface/pores is undesirable¹⁰⁸. However, production of clean biochar
585 is feasible and can be achieved by tuning a few key process parameters such as peak
586 temperature, feedstock type, residence time and carrier gas flow rate¹¹¹⁻¹¹³. It is important for
587 the biochar production process to be designed and operated in a way that yields biochar with
588 minimal content of these toxic compounds. Biochar production and characterisation should
589 be performed as per EBC and IBI biochar standard guidelines to ensure its safe use,
590 especially in water treatment^{114,115}. There is still a lack of research related to standardising
591 biochar production and hence there is an urgent need to have benchmarks and standards for
592 biochar materials, given the large spectrum of feedstock, production conditions and
593 properties of biochar¹¹⁶.

594 Carbon pricing is likely to become the norm in the coming years for effective climate change
595 mitigation. Carbon prices will need to be sufficiently high in order to restrict the global
596 warming to less than 2°C. Development of carbon-negative technologies such as biochar can
597 be expected to receive more attention in the future. Apart from having systematic life-cycle
598 assessments and techno-economic analysis specifically for BEBs, these analyses must also
599 account for carbon pricing in correctly estimating the economic and sustainable benefits of
600 biochar for energy and environmental applications, including water treatment. While there
601 are a few studies on biochar stability and aging, more studies should systematically
602 investigate the aging and stability of biochar to truly account for the carbon sequestration
603 potential of different types of biochar¹¹⁷⁻¹¹⁹.

604 **7. Conclusions**

605 This review highlights the potential of Biologically-Enhanced Biochar as a sustainable, cost-
606 effective and efficient biological water treatment technology in line with the zero waste
607 concept. Even though BEBs can effectively mediate biodegradation via several complex
608 mechanisms, there is no consensus on why and how certain types of BEBs work better for
609 certain process conditions and contaminant types. A bottom-up approach incorporating
610 research inputs from several disciplines is necessary to fully decode the complex and
611 dynamic biochar-microbial interactions in water treatment. We need to understand how
612 changes in biochar properties and process conditions can bring variations in biofilm microbial
613 diversity. It is also imperative to understand how these changes would reflect in the
614 efficiency and mechanisms of contaminant removal. Research focusing on molecular level
615 interactions of BEB, such as transport phenomena (over several diffusion layers),
616 biodegradation kinetics, biodegradation mechanisms (adsorption, biosorption,
617 biodegradation, bio-electrochemical interaction) and biofilm metabolic potential are
618 necessary. We need more research focusing on toxicity studies of biochar and biodegradation
619 products from BEBs to adopt sequential biochar systems in a safe and cost-effective manner.
620 Mathematical models and statistical tools to support experimental data and accurately
621 correlate the properties and control parameters of an efficient BEB filter are necessary.

622 If correctly designed, Biologically-Enhanced Biochar filters provide a sustainable solution for
623 water treatment, especially in LMICs with additional economic and societal benefits to local
624 communities. The utilisation of BEB can clearly contribute to meeting many of the United
625 Nations Sustainable Development Goals (SDG) by 2030 and especially SDG 6 (Clean water
626 and Sanitation) and SDG 7 (Affordable and Clean energy).

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632 **9. References**

- 633 1. Progress on Drinking Water, Sanitation and Hygiene: 2017 Update and SDG Baselines
634 (World Health Organization (WHO) and United Nations Children's Fund (UNICEF),
635 2017), <https://www.who.int/news-room/fact-sheets/detail/drinking-water>
- 636 2. Sato, T., Qadir, M., Yamamoto, S., Endo, T. & Zahoor, A. Global, regional, and
637 country level need for data on wastewater generation, treatment, and use. *Agric. Water*
638 *Manag.* **130**, 1–13 (2013).
- 639 3. Alvarino, T., Suarez, S., Lema, J. & Omil, F. Understanding the sorption and
640 biotransformation of organic micropollutants in innovative biological wastewater
641 treatment technologies. *Sci. Total Environ.* **615**, 297–306 (2018).
- 642 4. Woolf, D., Amonette, J. E., Street-Perrott, F. A., Lehmann, J. & Joseph, S. J N.
643 communications. Sustainable biochar to mitigate global climate change. **1**, 56 (2010).
- 644 5. Çeçen, F. & Aktas, Ö. Activated carbon for water and wastewater treatment:
645 integration of adsorption and biological treatment. (John Wiley & Sons, 2011).
- 646 6. Qambrani, N. A., Rahman, M. M., Won, S., Shim, S. & Ra, C. Biochar properties and
647 eco-friendly applications for climate change mitigation, waste management, and
648 wastewater treatment: A review. **79**, 255–273 (2017).
- 649 7. Meyer, S., Glaser, B. & Quicker, P. Technical, economical, and climate-related aspects

- 650 of biochar production technologies: a literature review. *Env. Sci Technol* **45**, 9473–
651 9483 (2011).
- 652 8. Liu, W.-J., Jiang, H. & Yu, H.-Q. Emerging applications of biochar-based materials
653 for energy storage and conversion. *Energy Environ. Sci.* **12**, 1751–1779 (2019).
- 654 9. Li, J., Fan, J., Zhang, J., Hu, Z. & Liang, S. Preparation and evaluation of wetland
655 plant-based biochar for nitrogen removal enhancement in surface flow constructed
656 wetlands. *Environ. Sci. Pollut. Res.* **25**, 13929–13937 (2018).
- 657 10. Inyang, M. & Dickenson, E. The potential role of biochar in the removal of organic
658 and microbial contaminants from potable and reuse water: A review. *Chemosphere*
659 **134**, 232–240 (2015).
- 660 11. Berenguer, R. *et al.* Electroactive Biochar: Sustainable and Scalable Environmental
661 Applications of Microbial Electrochemical Technologies. (2019).
- 662 12. Wang, C. *et al.* Role of biochar in the granulation of anaerobic sludge and
663 improvement of electron transfer characteristics. *Bioresour. Technol.* **268**, 28–35
664 (2018).
- 665 13. Zhu, X., Chen, B., Zhu, L. & Xing, B. Effects and mechanisms of biochar-microbe
666 interactions in soil improvement and pollution remediation: A review. *Env. Pollut* **227**,
667 98–115 (2017).
- 668 14. Zhao, Z. *et al.* Potential enhancement of direct interspecies electron transfer for
669 syntrophic metabolism of propionate and butyrate with biochar in up-flow anaerobic
670 sludge blanket reactors. **209**, 148–156 (2016).
- 671 15. Henze, M., van Loosdrecht, M. C. M., Ekama, G. A. & Brdjanovic, D. *Biological*
672 *wastewater treatment*. (IWA publishing, 2008).

- 673 16. Servais, P., Billen, G. & Bouillot, P. Biological Colonization of Granular Activated
674 Carbon Filters in Drinking Water Treatment, *J. Environ. Engg.* **120**, **4**, 888–899 (1994).
- 675 17. Jin, P., Jin, X., Wang, X., Feng, Y. & C, X. Biological Activated Carbon Treatment
676 Process for Advanced Water and Wastewater Treatment. (2013) in *Biomass Now:*
677 *Cultivation and Utilisation*, doi:10.5772/52021
- 678 18. Simpson, D. R. Biofilm processes in biologically active carbon water purification.
679 *Water Res* **42**, 2839–2848 (2008).
- 680 19. Wurzer Sohi, S. and Masek, O, C. Synergies in sequential biochar systems. in
681 *Advanced Carbon Materials from Biomass: an Overview* (ed. Manya, J. J.) 147–159
682 (Zenodo, 2019). doi:DOI: 10.5281/zenodo.3233732
- 683 20. Guardabassi, L., Lo Fo Wong, D. M. A. & Dalsgaard, A. The effects of tertiary
684 wastewater treatment on the prevalence of antimicrobial resistant bacteria. *Water Res.*
685 **36**, 1955–1964 (2002).
- 686 21. Bolisetty, S., Peydayesh, M. & Mezzenga, R. Sustainable technologies for water
687 purification from heavy metals: review and analysis. *Chem. Soc. Rev.* **48**, 463–487
688 (2019).
- 689 22. Korotta-Gamage, S. M. & Sathasivan, A. A review: Potential and challenges of
690 biologically activated carbon to remove natural organic matter in drinking water
691 purification process. *Chemosphere* **167**, 120–138 (2017).
- 692 23. Lohwacharin, J., Phetrak, A., Takizawa, S., Kanisawa, Y. & Okabe, S. Bacterial
693 growth during the start-up period of pilot-scale biological activated carbon filters:
694 Effects of residual ozone and chlorine and backwash intervals. *Process Biochem.* **50**,
695 1640–1647 (2015).

- 696 24. Bouabidi, Z. B., El-Naas, M. H. & Zhang, Z. Immobilization of microbial cells for the
697 biotreatment of wastewater: A review. *Environ. Chem. Lett.* **17**, 241–257 (2019).
- 698 25. Sharma, A., Jamali, H., Vaishnav, A., Giri, B. S. & Srivastava, A. K. Chapter 15 -
699 Microbial biofilm: An advanced eco-friendly approach for bioremediation. in *New and*
700 *Future Developments in Microbial Biotechnology and Bioengineering: Microbial*
701 *Biofilms* (eds. Yadav, M. K. & Singh, B. P.) 205–219 (Elsevier, 2020).
- 702 26. Lin, C.-K., Tsai, T.-Y., Liu, J.-C. & Chen, M.-C. Enhanced biodegradation of
703 petrochemical wastewater using ozonation and bac advanced treatment system. *Water*
704 *Res.* **35**, 699–704 (2001).
- 705 27. Smolin, S., Kozyatnyk, I. & Klymenko, N. New approach for the assessment of the
706 contribution of adsorption, biodegradation and self-bioregeneration in the dynamic
707 process of biologically active carbon functioning. *Chemosphere* **248**, 126022 (2020).
- 708 28. Logan, B. E., Rossi, R., Ragab, A. & Saikaly, P. E. Electroactive microorganisms in
709 bioelectrochemical systems. *Nat Rev Microbiol* **17**, 307–319 (2019).
- 710 29. Gwenz, W., Chaukura, N., Noubactep, C. & Mukome, F. N. D. Biochar-based water
711 treatment systems as a potential low-cost and sustainable technology for clean water
712 provision. *J Env. Manag.* **197**, 732–749 (2017).
- 713 30. Inyang, M. I. *et al.* A review of biochar as a low-cost adsorbent for aqueous heavy
714 metal removal. **46**, 406–433 (2016).
- 715 31. Sizmur, T., Fresno, T., Akgul, G., Frost, H. & Moreno-Jimenez, E. Biochar
716 modification to enhance sorption of inorganics from water. *Bioresour Technol* **246**,
717 34–47 (2017).
- 718 32. Xiang, W. *et al.* Biochar technology in wastewater treatment: A critical review.

- 719 *Chemosphere* **252**, 126539 (2020).
- 720 33. Hiew, B. Y. Z., Lee, L. Y., Thangalazhy-Gopakumar, S. & Gan, S. Biosorption. in
721 *Bioprocess Engineering* 143–164 (CRC Press, 2019).
- 722 34. Frankel, M. L. *et al.* Removal and biodegradation of naphthenic acids by biochar and
723 attached environmental biofilms in the presence of co-contaminating metals. *Bioresour*
724 *Technol* **216**, 352–361 (2016).
- 725 35. Kurniawan, A. %J J. of E. E. & Technology, S. Biofilm matrices as biomonitoring
726 agent and biosorbent for Cr (VI) pollution in aquatic ecosystems. **5**, 61–67 (2019).
- 727 36. Zhu, N., Zhang, J., Tang, J., Zhu, Y. & Wu, Y. Arsenic removal by periphytic biofilm
728 and its application combined with biochar. *Bioresour Technol* **248**, 49–55 (2018).
- 729 37. Dalahmeh, S., Ahrens, L., Gros, M., Wiberg, K. & Pell, M. Potential of biochar filters
730 for onsite sewage treatment: Adsorption and biological degradation of pharmaceuticals
731 in laboratory filters with active, inactive and no biofilm. *Sci. Total Environ.* **612**, 192–
732 201 (2018).
- 733 38. Dalahmeh, S. S., Alziq, N. & Ahrens, L. Potential of biochar filters for onsite
734 wastewater treatment: Effects of active and inactive biofilms on adsorption of per- and
735 polyfluoroalkyl substances in laboratory column experiments. *Env. Pollut* **247**, 155–
736 164 (2019).
- 737 39. Afrooz, A. R. & Boehm, A. B. Escherichia coli Removal in Biochar-Modified
738 Biofilters: Effects of Biofilm. *PLoS One* **11**, e0167489 (2016).
- 739 40. Mohanty, S. K. & Boehm, A. B. Escherichia coli Removal in Biochar-Augmented
740 Biofilter: Effect of Infiltration Rate, Initial Bacterial Concentration, Biochar Particle
741 Size, and Presence of Compost. *Environ. Sci. Technol.* **48**, 11535–11542 (2014).

- 742 41. Sasidharan, S. *et al.* Transport and retention of bacteria and viruses in biochar-
743 amended sand. **548**, 100–109 (2016).
- 744 42. Boehm, A. B. *et al.* Biochar-augmented biofilters to improve pollutant removal from
745 stormwater—can they improve receiving water quality? (2020).
- 746 43. Afrooz, A. R. M. N., Pitol, A. K., Kitt, D. & Boehm, A. B. Role of microbial cell
747 properties on bacterial pathogen and coliphage removal in biochar-modified
748 stormwater biofilters. *Environ. Sci. Water Res. Technol.* **4**, 2160–2169 (2018).
- 749 44. Edeh, I. G., Mašek, O. & Buss, W. A meta-analysis on biochar’s effects on soil water
750 properties – New insights and future research challenges. *Sci. Total Environ.* **714**,
751 136857 (2020).
- 752 45. Afrooz, A. R. M. N., Pitol, A. K., Kitt, D., Boehm, A. B. Role of microbial cell
753 properties on bacterial pathogen and coliphage removal in biochar-modified
754 stormwater biofilters. *Environ. Sci.: Water Res. Technol.*, 2018,**4**, 2160-2169
- 755 46. Roden, E. E. *et al.* Extracellular electron transfer through microbial reduction of solid-
756 phase humic substances. *Nat. Geosci.* **3**, 417–421 (2010).
- 757 47. Shi, L. *et al.* Extracellular electron transfer mechanisms between microorganisms and
758 minerals. *Nat. Rev. Microbiol.* **14**, 651–662 (2016).
- 759 48. Klupfel, L., Keiluweit, M., Kleber, M. & Sander, M. Redox properties of plant
760 biomass-derived black carbon (biochar). *Env. Sci Technol* **48**, 5601–5611 (2014).
- 761 49. Schievano, A. *et al.* Electroactive Biochar for Large-Scale Environmental Applications
762 of Microbial Electrochemistry. *ACS Sustain. Chem. Eng.* **7**, 18198–18212 (2019).
- 763 50. García-Rodríguez, J. P., Amezquita-García, H. J., Escamilla-Alvarado, C., Rangel-

- 764 Mendez, J. R. & Gutiérrez-García, K. Biofilm microbial composition changes due to
765 different surface chemical modifications of activated carbon cloths in the
766 biotransformation of 4-nitrophenol. *Biodegradation* **30**, 401–413 (2019).
- 767 51. Korth, B., Rosa, L. F. M., Harnisch, F. & Picioreanu, C. A framework for modeling
768 electroactive microbial biofilms performing direct electron transfer.
769 *Bioelectrochemistry* **106**, 194–206 (2015).
- 770 52. Koch, C. & Harnisch, F. What Is the Essence of Microbial Electroactivity? *Front.*
771 *Microbiol.* **7**, (2016).
- 772 53. Prado, A., Berenguer, R. & Esteve-Núñez, A. Electroactive biochar outperforms
773 highly conductive carbon materials for biodegrading pollutants by enhancing microbial
774 extracellular electron transfer. *Carbon N. Y.* **146**, 597–609 (2019).
- 775 54. Klüpfel, L., Piepenbrock, A., Kappler, A. & Sander, M. Humic substances as fully
776 regenerable electron acceptors in recurrently anoxic environments. *Nat. Geosci.* **7**,
777 195–200 (2014).
- 778 55. Ramírez-Vargas, C. A. *et al.* Microbial Electrochemical Technologies for Wastewater
779 Treatment: Principles and Evolution from Microbial Fuel Cells to Bioelectrochemical-
780 Based Constructed Wetlands. **10**, 1128 (2018).
- 781 56. Logan, B. E. & Rabaey, K. Conversion of Wastes into Bioelectricity and Chemicals by
782 Using Microbial Electrochemical Technologies. **337**, 686–690 (2012).
- 783 57. Semenc, L., Aracic, S., Mathews, E. R., Franks, Electron Transfer Between Bacteria
784 and Electrodes in Functional Electrodes for Enzymatic and Microbial Electrochemical
785 Systems, (CNRS, France), (2017).
- 786 58. Gorovtsov, A. V *et al.* The mechanisms of biochar interactions with microorganisms in

- 787 soil. *Env. Geochem Heal.* 42, 2495–2518, (2020)
- 788 59. Lovley, D. R. Live wires: direct extracellular electron exchange for bioenergy and the
789 bioremediation of energy-related contamination. *Energy Environ. Sci.* 4, 4896 (2011).
- 790 60. Chen, S. *et al.* Promoting interspecies electron transfer with biochar. *Sci Rep* 4, 5019
791 (2014).
- 792 61. Yuan, Y. *et al.* Applications of biochar in redox-mediated reactions. *Bioresour*
793 *Technol* 246, 271–281 (2017).
- 794 62. Sun, T. *et al.* Rapid electron transfer by the carbon matrix in natural pyrogenic carbon.
795 *Nat Commun* 8, 14873 (2017).
- 796 63. Barua, S. & Dhar, B. R. J B. technology. Advances towards understanding and
797 engineering direct interspecies electron transfer in anaerobic digestion. *Bioresour*
798 *Technol* 244, 698–707 (2017).
- 799 64. Yapsakli, K. & Çeçen, F. Effect of type of granular activated carbon on DOC
800 biodegradation in biological activated carbon filters. *Process Biochem.* 45, 355–362
801 (2010).
- 802 65. Lu, Z. *et al.* Effect of granular activated carbon pore-size distribution on biological
803 activated carbon filter performance. *Water Res.* 177, 115768 (2020).
- 804 66. Fundneider, T., Acevedo Alonso, V., Wick, A., Albrecht, D. & Lackner, S.
805 Implications of biological activated carbon filters for micropollutant removal in
806 wastewater treatment. *Water Res.* 189, 116588 (2021).
- 807 67. Maxence Carrel, Verónica L. Morales, Mario A. Beltran, Nicolas Derlon, Rolf
808 Kaufmann, Eberhard Morgenroth, Markus Holzner, Biofilms in 3D porous media:

- 809 Delineating the influence of the pore network geometry, flow and mass transfer on
810 biofilm development, *Water Res.* **134**, 280-291 (2018)
- 811 68. Gerlach R, Cunningham AB, "Influence of biofilms on porous media hydrodynamics,"
812 In: Porous Media: Applications in Biological Systems and Biotechnology, ed. Vafai K,
813 CRC Press Taylor Francis Group 2010 pp 173-230.
- 814 69. Oh, S., Hammes, F. & Liu, W.-T. Metagenomic characterization of biofilter microbial
815 communities in a full-scale drinking water treatment plant. *Water Res.* **128**, 278–285
816 (2018).
- 817 70. Wu, L. *et al.* Global diversity and biogeography of bacterial communities in
818 wastewater treatment plants. *Nat. Microbiol.* **4**, 1183–1195 (2019).
- 819 71. Hill, R. A. *et al.* Effect of Biochar on Microbial Growth: A Metabolomics and
820 Bacteriological Investigation in *E. coli*. *Environ. Sci. Technol.* **53**, 2635–2646 (2019).
- 821 72. Dalahmeh, S. S. *et al.* Dynamics and functions of bacterial communities in bark,
822 charcoal and sand filters treating greywater. *Water Res.* **54**, 21–32 (2014).
- 823 73. Hagemann, N. *et al.* Activated Carbon, Biochar and Charcoal: Linkages and Synergies
824 across Pyrogenic Carbon's ABCs. *Water* **10**, 182 (2018).
- 825 74. Activated Carbon Market Analysis Size, Share, Growth, Trends and Segment
826 Forecasts To 2020: Grand View Research. *M2 Communications* Activated Carbon
827 Market Analysis Size, Share, Growth, Trends and Segment Forecasts To 2020: Grand
828 View Research, Report ID: GVR-4-68038-011-8, Report ID: 978-1-68038-073-6,
829 <https://www.grandviewresearch.com/industry-analysis/activated-carbon-market>,
830 <https://www.grandviewresearch.com/industry-analysis/coal-based-activated-carbon-market> (2015).
- 831 75. Alhashimi, H. A. & Aktas, C. B. Life cycle environmental and economic performance

- 832 of biochar compared with activated carbon: A meta-analysis. *Resour. Conserv. Recycl.*
833 **118**, 13–26 (2017).
- 834 76. Thompson, K. A. *et al.* Environmental Comparison of Biochar and Activated Carbon
835 for Tertiary Wastewater Treatment. *Env. Sci Technol* **50**, 11253–11262 (2016).
- 836 77. El Gamal, M., Mousa, H. A., El-Naas, M. H., Zacharia, R. & Judd, S. Bio-regeneration
837 of activated carbon: A comprehensive review. *Sep. Purif. Technol.* **197**, 345–359
838 (2018).
- 839 78. Gabarrell, X. *et al.* A comparative life cycle assessment of two treatment technologies
840 for the Grey Lanaset G textile dye: biodegradation by *Trametes versicolor* and granular
841 activated carbon adsorption. **17**, 613–624 (2012).
- 842 79. Bayer, P., Heuer, E., Karl, U. & Finkel, M. Economical and ecological comparison of
843 granular activated carbon (GAC) adsorber refill strategies. *Water Res.* **39**, 1719–1728
844 (2005).
- 845 80. Chen, J. *et al.* A review of biomass burning: Emissions and impacts on air quality,
846 health and climate in China. *Sci. Total Environ.* **579**, 1000–1034 (2017).
- 847 81. Goswami, S. B., Mondal, R. & Mandi, S. K. Crop residue management options in
848 rice–rice system: a review. *Arch. Agron. Soil Sci.* 1–17 (2019).
- 849 82. Mathur, R. & Srivastava, V. K. Crop Residue Burning: Effects on Environment. in
850 *Greenhouse Gas Emissions: Challenges, Technologies and Solutions* (eds. Shurpali,
851 N., Agarwal, A. K. & Srivastava, V. K.) 127–140 (Springer Singapore, 2019).
- 852 83. Ravindra, K., Singh, T. & Mor, S. Emissions of air pollutants from primary crop
853 residue burning in India and their mitigation strategies for cleaner emissions. *J. Clean.*
854 *Prod.* **208**, 261–273 (2019).

- 855 84. Smebye, A. B., Sparrevik, M., Schmidt, H. P. & Cornelissen, G. Life-cycle assessment
856 of biochar production systems in tropical rural areas: Comparing flame curtain kilns to
857 other production methods. *Biomass and Bioenergy* **101**, 35–43 (2017).
- 858 85. Pandit, N. R., Mulder, J., Hale, S. E., Schmidt, H. P. & Cornelissen, G. Biochar from
859 ‘Kon Tiki’ flame curtain and other kilns: Effects of nutrient enrichment and kiln type
860 on crop yield and soil chemistry. *PLoS One* **12**, 1–18 (2017).
- 861 86. Yoder, J., Galinato, S., Granatstein, D. & Garcia-Pérez, M. Economic tradeoff between
862 biochar and bio-oil production via pyrolysis. *Biomass and Bioenergy* **35**, 1851–1862
863 (2011).
- 864 87. Van Laer, T. *et al.* Legal constraints and opportunities for biochar: a case analysis of
865 EU law. *GCB Bioenergy*, 7, 14–24 (2015).
- 866 88. Kozyatnyk, I., Yacout, D. M. M., Van Caneghem, J. & Jansson, S. Comparative
867 environmental assessment of end-of-life carbonaceous water treatment adsorbents.
868 *Bioresour. Technol.* **302**, 122866 (2020).
- 869 89. Boateng, A. A., Garcia-Perez, M., Masek, O., Brown, R. & del Campo, B. Biochar
870 production technology. *Biochar Environ. Manag. Technol.* 63–109 (2015).
- 871 90. Stavropoulos, G. G. & Zabaniotou, A. A. Minimizing activated carbons production
872 cost. *Fuel Process. Technol.* **90**, 952–957 (2009).
- 873 91. Wang, D., Jiang, P., Zhang, H. & Yuan, W. Biochar production and applications in
874 agro and forestry systems: A review. *Sci. Total Environ.* **723**, 137775 (2020).
- 875 92. Ahmed, M. B., Zhou, J. L., Ngo, H. H. & Guo, W. Adsorptive removal of antibiotics
876 from water and wastewater: Progress and challenges. *Sci. Total Environ.* **532**, 112–126
877 (2015).

- 878 93. Novak, J. *et al.* Biochars multifunctional role as a novel technology in the agricultural,
879 environmental, and industrial sectors. *Chemosphere* **142**, 1–3 (2016).
- 880 94. Shepherd, J. G., Sohi, S. P. & Heal, K. V. Optimising the recovery and re-use of
881 phosphorus from wastewater effluent for sustainable fertiliser development. *Water*
882 *Res.* **94**, 155–165 (2016).
- 883 95. Qian, T., Lu, D., Soh, Y. N. A., Webster, R. D. & Zhou, Y. Biotransformation of
884 phosphorus in enhanced biological phosphorus removal sludge biochar. *Water Res.*
885 **169**, 115255 (2020).
- 886 96. Yang, H. *et al.* Utilization of biochar for resource recovery from water: A review.
887 *Chem. Eng. J.* **397**, 125502 (2020).
- 888 97. Indren, M., Birzer, C. H., Kidd, S. P., Hall, T. & Medwell, P. R. Effects of biochar
889 parent material and microbial pre-loading in biochar-amended high-solids anaerobic
890 digestion. *Bioresour. Technol* **298**, 122457 (2020).
- 891 98. Jang, H. M., Choi, Y.-K. & Kan, E. Effects of dairy manure-derived biochar on
892 psychrophilic, mesophilic and thermophilic anaerobic digestions of dairy manure.
893 *Bioresour. Technol* **250**, 927–931 (2018).
- 894 99. Cheong, J. C. *et al.* Closing the food waste loop: Food waste anaerobic digestate as
895 fertilizer for the cultivation of the leafy vegetable, xiao bai cai (*Brassica rapa*). *Sci.*
896 *Total Environ.* **715**, 136789 (2020).
- 897 100. McDowell, D. *et al.* Recycling nutrients from anaerobic digestates for the cultivation
898 of *Phaeodactylum tricornutum*: A feasibility study. *Algal Res.* **48**, 101893 (2020).
- 899 101. Huggins, T. M., Latorre, A., Biffinger, J. C. & Ren. Biochar based microbial fuel cell
900 for enhanced wastewater treatment and nutrient recovery. *Sustainability* **8**, 169 (2016).

- 901 102. Bamdad, H., Hawboldt, K. & MacQuarrie, S. A review on common adsorbents for acid
902 gases removal: Focus on biochar. *Renew. Sustain. Energy Rev.* **81**, 1705–1720 (2018).
- 903 103. Choudhury, A. & Lansing, S. Biochar addition with Fe-impregnation to reduce H₂S
904 production from anaerobic digestion. *Bioresour. Technol* 123121 (2020).
- 905 104. Sethupathi, S. *et al.* Biochars as potential adsorbents of CH₄, CO₂ and H₂S. **9**, 121
906 (2017).
- 907 105. Das, J. *et al.* Performance of a compost and biochar packed biofilter for gas-phase
908 hydrogen sulfide removal. **273**, 581–591 (2019).
- 909 106. Nemani, V. A., McKie, M. J., Taylor-Edmonds, L. & Andrews, R. C. Impact of
910 biofilter operation on microbial community structure and performance. *J. Water*
911 *Process Eng.* **24**, 35–41 (2018).
- 912 107. Ibn Abdul Hamid, K., Sanciolo, P., Gray, S., Duke, M. & Muthukumaran, S.
913 Comparison of the effects of ozone, biological activated carbon (BAC) filtration and
914 combined ozone-BAC pre-treatments on the microfiltration of secondary effluent. *Sep.*
915 *Purif. Technol.* **215**, 308–316 (2019).
- 916 108. Li, W.-G., Qin, W., Song, Y., Zheng, Z.-J. & Lv, L.-Y. Impact of ozonation and
917 biologically enhanced activated carbon filtration on the composition of micropollutants
918 in drinking water. *Environ. Sci. Pollut. Res.* **26**, 33927–33935 (2019).
- 919 109. Boon, N., Pycke, B. F. G., Marzorati, M. & Hammes, F. Nutrient gradients in a
920 granular activated carbon biofilter drives bacterial community organization and
921 dynamics. *Water Res.* **45**, 6355–6361 (2011).
- 922 110. Buss, W., Masek, O., Graham, M. & Wust, D. Inherent organic compounds in biochar-
923 -Their content, composition and potential toxic effects. *J Env. Manag.* **156**, 150–157

- 924 (2015).
- 925 111. Manyà, J. J. *Advanced Carbon Materials from Biomass: an Overview*. (Zenodo, 2019).
- 926 112. Manyà, J. J. Pyrolysis for biochar purposes: a review to establish current knowledge
927 gaps and research needs. *Env. Sci Technol* **46**, 7939–7954 (2012).
- 928 113. Buss, W., Graham, M. C., MacKinnon, G., Mašek, O.. Strategies for producing
929 biochars with minimum PAH contamination. *J. Anal. Appl. Pyrolysis* **119**, 24–30
930 (2016).
- 931 114. European Biochar Foundation (EBC). Guidelines for a Sustainable Production of
932 Biochar. *Eur. Biochar Found.* 1–22 (2016). doi:10.13140/RG.2.1.4658.7043
- 933 115. IBI biochar standards, <https://biochar-international.org/characterizationstandard/>
- 934 116. Mašek, O. *et al.* Consistency of biochar properties over time and production scales: A
935 characterisation of standard materials. *J. Anal. Appl. Pyrolysis* **132**, 200–210 (2018).
- 936 117. Kim, H.-B., Kim, J.-G., Kim, T., Alessi, D. S. & Baek, K. Interaction of biochar
937 stability and abiotic aging: Influences of pyrolysis reaction medium and temperature.
938 *Chem. Eng. J.* **411**, 128441 (2021).
- 939 118. Rathnayake, D. *et al.* How to trace back an unknown production temperature of
940 biochar from chemical characterization methods in a feedstock independent way. *J.*
941 *Anal. Appl. Pyrolysis* **151**, 104926 (2020).
- 942 119. Crombie, K., Mašek, O., Sohi, S. P., Brownsort, P. & Cross, A. The effect of pyrolysis
943 conditions on biochar stability as determined by three methods. *GCB Bioenergy* **5**,
944 122–131 (2013).

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Supporting Information

New directions and challenges in engineering Biologically-Enhanced Biochar for biological water treatment

Paper selection for review

Google Scholar Advanced search engine was used to find relevant papers in the subject. Close to 300 papers were screened. When we were faced with choosing papers from multiple options (several papers in the same field), criteria was to choose the paper that best described and discussed the specific point in question. In doing this, care has also been taken to come up with the most recent and highly cited literature that discussed the most recent developments in this field. Journal matrices and following research groups/authors who work in this specific field were very helpful. Missing gaps in literature and the pressing issues associated with these gaps were compiled during this process. This was done using several mind maps and short topic proposals. This was helpful to finalise this specific topic that we chose to review. We were able to find more papers from the reference lists of papers selected in the first screening. The initial screening, paper selection, the specific topic selection, and finally the critical review of relevant literature was all done in a span of 6-8 months, with inputs and discussion from all the contributing authors. The keywords below were rearranged in various combinations, including suggestions from Google scholar after each search, to compile more specific information.

- Water treatment steps
- Biological water treatment
- Activated carbon production
- Biochar production
- Biological activated carbon
- Biochar biofilm interactions

- Microbial biofilms for water treatment
- Biochar characterisation
- Electrical and electrochemical properties of biochar
- Biofilm properties and characterisation
- Modelling biochar properties
- Microbial biofilm modelling
- Biochar Biofilm interactions
- Life cycle analysis of biochar and activated carbon for water treatment
- Environmental comparison of biochar and activated carbon
- Biochar for sustainability
- Nutrient recovery from water treatment
- Biochar for nutrient recovery
- Multifunctional biochar
- Biofilm ecology and biological water treatment
- Biofilm composition and biological activated carbon